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Draft Genome Sequence of *Aneurinibacillus* sp. Strain BA2021, Isolated as a Contaminant of a Laboratory-Cultivated Predatory Myxobacterium

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ABSTRACT During laboratory cultivation of the myxobacterium *Archangium violaceum* strain Cb vi76, a reoccurring contaminant was isolated and sequenced. Comparative taxonomic analysis of the draft genome suggested the contaminant to be a novel species, currently designated *Aneurinibacillus* sp. strain BA2021, from the genus *Aneurinibacillus*, members of which are considered promising biocontrol agents.

The discovery of novel motile spore-forming bacteria of the genus *Aneurinibacillus* offers a promising advancement in the development of sustainable biocontrol agents (1, 2). Significantly, *Aneurinibacillus migulanus* has been reported to produce the cyclic peptide gramicidin S with activities against a wide range of bacteria and fungi (2, 3). Here, we report a draft genome sequence for *Aneurinibacillus* sp. strain BA2021, which was isolated as a reoccurring contaminant during the laboratory cultivation of a stock of a predatory myxobacterium, *Archangium violaceum* strain Cb vi76, obtained from the Leibniz Institute DSMZ (German Collection of Microorganisms and Cell Cultures GmbH).

Aneurinibacillus sp. strain BA2021 cells were passaged from the originally observed coculture with A. violaceum and initially cultivated as a monoculture on VY/2 agar (5 g/liter baker's yeast, 1.36 g/liter $CaCl_{2r}$, 0.5 mg/liter vitamin B_{12r} and 15 g/liter agar [pH 7.2]) plates for 8 days at 30°C, transferred to CTTYE broth (10 g/liter Casitone, 5 g/liter yeast extract, 1 mM KH₂PO₄, 8 mM MgSO₄, 10 mM Tris-HCl [pH 7.6]), and incubated as described above. Genomic DNA was isolated from harvested cells using a NucleoBond high-molecular-weight DNA purification kit (Macherey-Nagel), with the initial DNA quantified using the Qubit double-stranded DNA (dsDNA) high-sensitivity (HS) assay kit (Life Technologies) and evaluated using a NanoDrop 2000 spectrophotometer (Thermo Fisher Scientific). Subsequent sample preparation and sequencing were performed at MR DNA (Shallowater, TX). Initial sample sizes of 1 to 15 kb were determined by electrophoresis (E-Gel SizeSelect 2% agarose gel; Invitrogen) and sheared (Covaris g-TUBE) to provide DNA fragments ranging from 6 to 10 kb, which were selected for subsequent sequencing by size using a Sage Science BluePippin automated size selection instrument. A SMRTbell Express template prep kit version 2.0 (Pacific Biosciences) was used to build the library according to the manufacturer's protocol. Each sample underwent DNA damage and end repair, as well as barcode adapter ligation. After the library construction, the final concentration of the library (8.18 ng/µL) was measured using the Qubit dsDNA HS assay kit (Thermo Fisher Scientific), and the average library size (7,964 bp) was determined using the Agilent 2100 Bioanalyzer. The resulting libraries were sequenced using the 10-h movie time on the PacBio Sequel system. De novo assembly of the genome into 3 contigs with protein-coding genes (N_{50} contig length, 4,200,366 bp) was accomplished using the SMRT Analysis Hierarchical Genome Assembly Process (HGAP; SMRTLink version 9.0.0), which includes preassembly, assembly, and consensus polishing. Results from the Falcon assembler paired with the Arrow polishing algorithm include a mean seed read length of

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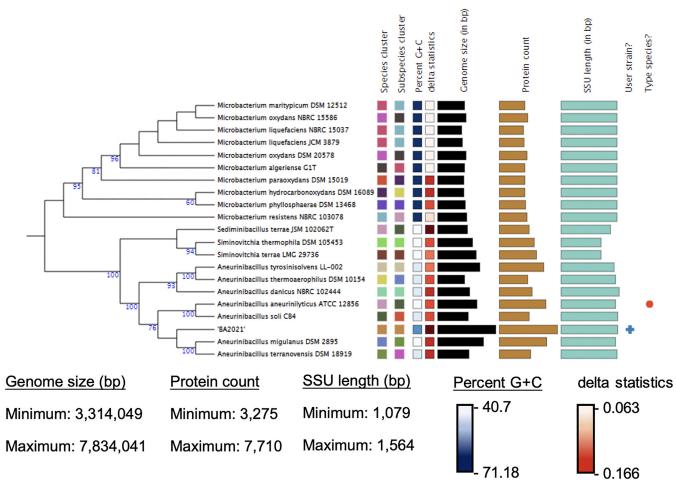


FIG 1 Maximum-likelihood phylogenetic tree suggesting Aneurinibacillus sp. strain BA2021 as a member of the genus Aneurinibacillus. Clustered species and subspecies are represented by the same colors. The following methods described were obtained from the server: The tree was rendered using the MASH algorithm to compare the draft genome sequence against all type strain genome sequences available in the TYGS database (6). Precise distances were calculated using the genome BLAST distance phylogeny approach (GBDP) with distance formula d_s to determine closely related type strains. The tree was rendered using FastME version 2.1.6.1 (7) with GBDP distances calculated from 16S rRNA gene sequences, and the numbers on the branches are GBDP pseudobootstrap support values of >60% from 100 replications, with an average branch support of 67.7% (8). SSU, small subunit.

10.3 kb (198,513 total reads), 91% realigned bases, and 87.7% mean realigned concordance. A mean coverage of $261 \times$ was achieved, and the lengths of the contigs were 4,200,366 bp, 3,559,306 bp, and 74,369 bp. Analysis of the genome completeness, performed using benchmarking universal single-copy orthologs (BUSCO) version 5.1.2 (4) with the "bacillales_odb10" data set with 447 BUSCOs, resulted in 97.3% complete and single-copy, 2% complete and duplicate-copy, 0.4% fragmented-copy, and 0.3% missing BUSCOs. Preliminary annotation, utilizing the Rapid Annotations using Subsystems Technology (RAST) server (5), showed that the draft genome sequence has 58.8% G+C content, 8,498 coding sequences, and a genome size of 7,834,041 bp, containing 216 identified RNAs. Subsequent annotation included with the deposited genome sequence was requested by the NCBI Prokaryotic Genome Annotation Pipeline (PGAP). Taxonomic identification of the genome sequence was performed with the Type (Strain) Genome Server (TYGS) (6) to determine the closest type strain genomes and generate a phylogenetic tree (Fig. 1).

Using antiSMASH version 6.0.0, six unique secondary metabolite biosynthetic pathways were identified, including pathways for two type III polyketides, one terpene, and two β -lactone-containing protease inhibitors (9). However, the biosynthetic pathway for the reported specialized metabolite from this genera, gramicidin S, was not observable in the antiSMASH analysis (10). Ultimately, we envision that the provided genome data will benefit investigations into the utility of members of the genus Aneurinibacillus as sustainable biocontrol agents.

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Data availability. This draft genome sequencing project has been deposited in DDBJ/ENA/GenBank under the accession number JAFJNS00000000. The version described in this paper is the first version, JAFJNS00000000.1. The raw data have been deposited under the BioProject accession number PRJNA698652 and in the Sequence Read Archive (SRA) under the accession number SRR13859926.

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REFERENCES

- 1. Chandel S, Allan EJ, Woodward S. 2010. Biological control of Fusarium oxysporum f.sp. lycopersici on tomato by Brevibacillus brevis. J Phytopathol 158:470-478. https://doi.org/10.1111/j.1439-0434.2009.01635.x.
- 2. Berditsch M, Afonin S, Ulrich AS. 2007. The ability of Aneurinibacillus migulanus (Bacillus brevis) to produce the antibiotic gramicidin S is correlated with phenotype variation. Appl Environ Microbiol 73:6620-6628. https://doi.org/10.1128/AEM.00881-07.
- 3. Alenezi FN, Rekik I, Bouket AC, Luptakova L, Weitz HJ, Rateb ME, Jaspars M, Woodward S, Belbahri L. 2017. Increased biological activity of Aneurinibacillus migulanus strains correlates with the production of new gramicidin secondary metabolites. Front Microbiol 8:517. https://doi.org/10 .3389/fmicb.2017.00517.
- 4. Seppey M, Manni M, Zdobnov EM. 2019. BUSCO: assessing genome assembly and annotation completeness. Methods Mol Biol 1962:227–245. https://doi.org/10.1007/978-1-4939-9173-0_14.
- 5. Overbeek R. Olson R. Pusch GD. Olsen GJ. Davis JJ. Disz T. Edwards RA. Gerdes S, Parrello B, Shukla M, Vonstein V, Wattam AR, Xia F, Stevens R. 2014. The SEED and the Rapid Annotation of microbial genomes using

- Subsystems Technology (RAST). Nucleic Acids Res 42:D206-D214. https:// doi.org/10.1093/nar/gkt1226.
- 6. Meier-Kolthoff JP, Göker M. 2019. TYGS is an automated high-throughput platform for state-of-the-art genome-based taxonomy. Nat Commun 10:2182. https://doi.org/10.1038/s41467-019-10210-3.
- 7. Lefort V, Desper R, Gascuel O. 2015. FastME 2.0: a comprehensive, accurate, and fast distance-based phylogeny inference program. Mol Biol Evol 32:2798-2800. https://doi.org/10.1093/molbev/msv150.
- 8. Farris JS. 1972. Estimating phylogenetic trees from distance matrices. Am Nat 106:645-668. https://doi.org/10.1086/282802.
- 9. Medema MH, Blin K, Cimermancic P, de Jager V, Zakrzewski P, Fischbach MA, Weber T, Takano E, Breitling R. 2011. antiSMASH: rapid identification, annotation and analysis of secondary metabolite biosynthesis gene clusters in bacterial and fungal genome sequences. Nucleic Acids Res 39: W339-W346. https://doi.org/10.1093/nar/gkr466.
- 10. Alenezi FN, Rekik I, Bełka M, Ibrahim AF, Luptakova L, Jaspars M, Woodward S, Belbahri L. 2016. Strain-level diversity of secondary metabolism in the biocontrol species Aneurinibacillus migulanus. Microbiol Res 182:116-124. https://doi.org/10.1016/j.micres.2015.10.007.