University of Mississippi

eGrove

Electronic Theses and Dissertations

Graduate School

1-1-2016

Does Blood Flow Restriction Applied Post High-Load Exercise Augment Skeletal Muscle Growth Following Eight Weeks of Training?

Scott Justin Dankel University of Mississippi

Follow this and additional works at: https://egrove.olemiss.edu/etd

Part of the Physiology Commons

Recommended Citation

Dankel, Scott Justin, "Does Blood Flow Restriction Applied Post High-Load Exercise Augment Skeletal Muscle Growth Following Eight Weeks of Training?" (2016). *Electronic Theses and Dissertations*. 1309. https://egrove.olemiss.edu/etd/1309

This Thesis is brought to you for free and open access by the Graduate School at eGrove. It has been accepted for inclusion in Electronic Theses and Dissertations by an authorized administrator of eGrove. For more information, please contact egrove@olemiss.edu.

DOES BLOOD FLOW RESTRICTION APPLIED POST HIGH-LOAD EXERCISE AUGMENT SKELETAL MUSCLE GROWTH FOLLOWING EIGHT WEEKS OF

TRAINING?

A Thesis presented in partial fulfillment of requirements for the degree of Master of Science In the Department of Health, Exercise Science and Recreation Management The University of Mississippi

by

Scott J. Dankel

May 2016

Copyright © Scott J. Dankel

May 2016

ALL RIGHTS RESERVED

ABSTRACT

The application of blood flow restriction during low load exercise has consistently been shown to augment muscle hypertrophy which has been attributed to metabolic accumulation. It remains unknown, however, whether metabolites can augment muscle growth independent of further mechanical tension, specifically when maintained post high-load training. Thirteen untrained individuals performed 24 training sessions. The control arm performed one set of elbow flexion (70% 1RM) exercise to volitional fatigue, while the experimental arm performed the same protocol immediately followed by 3 min of blood flow restriction (70% arterial occlusion). Both conditions completed the same volume (3687 vs. 3638 kg) of exercise. There was an interaction (p=0.031) demonstrating an attenuation of muscle growth at the 60% site in the experimental [pre: 3.1 (0.6), post: 3.1 (0.7) cm] vs. control [pre: 3.1 (0.7), post 3.3 (0.7) cm] condition. Muscle growth at the 50% site did not differ between the experimental [pre: 2.9 (0.6), post 2.9 (0.6) cm] and control [pre: 2.8 (0.7), post: 2.9 (0.6) cm] condition (p=0.31) nor did it differ at the 70% site [experimental pre: 3.3 (0.60), post 3.5 (0.7) cm; control pre: 3.4 (0.7), post 3.6(0.7) cm]. Although there were no differences at the group level, there were attenuations at the individual level. The number of measured sites displaying growth at or outside the error of the measurement was greater in the control (21) vs. experimental (10) condition. The application of blood flow restriction post high-load exercise did not augment, but appeared to attenuate muscle growth at the group and individual level. With regard to one-repetition maximum strength, increases were observed in both the control [pre: 13.5 (3.8), post: 16.3 (4.5) kg] and experimental [pre: 13.7 (4.1), post: 16.3 (4.6) kg] conditions with no differences between

conditions. No changes were observed for isometric or isokinetic strength for either the control or experimental conditions. These results unveil the possibilities that 1) metabolites do not have anabolic properties per se, and may be detrimental for muscle hypertrophy; 2) immediate post-exercise blood flow is important for muscle hypertrophy; and/or 3) metabolites have anabolic properties but this was masked by the restriction of blood flow.

LIST OF ABBREVIATIONS AND SYMBOLS

- 1RM One-repetition maximum
- Akt Protein kinase B
- AMPK AMP-activated protein kinase
- ANOVA Analysis of variance
- BFR Blood flow restriction
- EMG Electromyography
- IGF-1 Insulin like growth factor 1
- MAFbx Muscle atrophy F box
- MPB Muscle protein breakdown
- mRNA Messenger ribonucleic acid
- mTOR Mechanistic target of rapamycin
- mTORC1 Mechanistic target of rapamycin complex 1
- mTORC2 Mechanistic complex of rapamycin complex 2
- MURF1 Muscle ring finger-1
- TSC2 Tuberous sclerosis complex 2

ACKNOWLEDGEMENTS

I would like to first and foremost thank Dr. Jeremy Loenneke for not only helping with the countless hours that have gone into this thesis project, but for the guidance and education necessary to complete it in its entirety. I also want to thank all the member of our lab who have assisted in this thesis project including fellow graduate students Sam Buckner, Brittany Counts, Matt Jessee, Grant Mouser and Kevin Mattocks, and professors Dr. Takashi Abe and Dr. Gilberto Laurentino. The data collection alone for this study would not have been possible without their assistance. Finally, I would like to thank the BioLayne Foundation for funding this project and enabling us to provide compensation for the participants involved.

TABLE OF CONTENTS

ABSTRACT	ii
LIST OF ABBREVIATIONS	iv
ACKNOWLEDGEMENTS	V
LIST OF TABLES	vii
LIST OF FIGURES	viii
CHAPTER 1: INTRODUCTION	1
CHAPTER 2: LITERATURE REVIEW	7
CHAPTER 3: METHODS	
CHAPTER 4: RESULTS AND DISCUSSION	23
CHAPTER 5: CONCLUSION	46
REFERENCES	49
VITA	

LIST OF TABLES

1. Individual Demographics	23
2. Muscle Thickness	25
3. Meaningful Increases in Muscle Thickness	26
4. Sex Differences in Muscle Thickness	32
5. One-Repetition Maximum (1RM) Strength	34
6. Sex Differences in One-Repetition Maximum (1RM) Strength	36
7. Isometric and Isokinetic Strength	37
8. Exercise Volume	40
9. Repetitions	41
10. Arm Circumference	42
11. Borg (CR10+) Ratings of Discomfort	44

LIST OF FIGURES

1. Muscle Thickness	26
2. Within Subject Differences in Muscle Thickness Between Conditions	27
3. Individual Changes in Muscle Thickness	28
4. Sex Differences in Muscle Thickness	31
5. One-Repetition Maximum (1RM) Strength	33
6. Individual Changes in One-Repetition Maximum (1RM) Strength	34
7. Sex Differences in One-Repetition Maximum (1RM) Strength	
8. Isometric and Isokinetic Strength	38
9. Individual Changes in Isometric and Isokinetic Strength	39
10. Exercise Volume	40
11. Total Repetitions	41
12. Arm Circumference	43
13. Borg (CR10+) Ratings of Discomfort	44

CHAPTER 1: INTRODUCTION

An increase in the size of a muscle fiber is known as muscle hypertrophy and is known to occur after repetitive bouts of resistance training in humans. Muscle hypertrophy involves a variety of complex cellular and molecular mechanisms responsible for the formation of new proteins (90). When the number of proteins synthesized is greater than the number of proteins degraded, a positive net protein balance occurs, causing an increase in muscle size (78).

It was originally thought that heavier loads were necessary to produce muscle hypertrophy (9); a notion further supported by the American College of Sports Medicine (3). However, it has since been shown that muscle hypertrophy can occur through the lifting of lighter loads as long as the exercise is performed to volitional fatigue (66). Furthermore, blood flow restriction training (BFR) has been shown to increase muscle size through the lifting of lighter loads (20-30% 1RM) without the need to exercise to volitional fatigue (54). The increases in muscle size seen from BFR are comparable to that of high load training (43, 55), and provide insight that there may be a number of mechanisms responsible for muscle hypertrophy.

The primary mechanism responsible for muscle hypertrophy during high load training is thought to be mechanical tension. Since BFR training is performed with lighter loads that otherwise do not result in increased muscle size (101), it is unlikely mechanical tension alone is sufficient to stimulate robust muscular hypertrophy. Therefore, increases in muscle size from BFR may be reliant on alternative mechanisms to elicit growth in the absence of high levels of mechanical tension. The application of BFR has not been shown to provide any further muscle adaptation when combined with high load training (44), but this has only been implemented intermittently due to high levels of participant discomfort (44). This provides some insight that the BFR stimulus may only augment the hypertrophic response when insufficient mechanical tension is present.

It is currently thought that a buildup of metabolites causes an increase in systemic hormones, growth factors, activation of higher threshold motor units (53) and cellular swelling (48). These mechanisms have been hypothesized to occur during commonly implemented BFR protocols and therefore may be acting in conjunction with mechanical tension to stimulate muscle growth. In contrast, applying a restrictive stimulus post exercise would allow for a prolonged period of cell swelling and metabolic accumulation without any additional muscle contraction or mechanical tension. However, the hypertrophic effects of prolonged cell swelling and metabolic accumulation at the conclusion of exercise, but without further mechanical tension, remains to be tested. The aforementioned BFR protocols used in conjunction with low load resistance training have not been designed to differentiate between the potential additive effects of these alternative hypertrophic mechanisms. BFR applied to immobilized limbs has been shown to attenuate disuse atrophy (11) supporting its effectiveness independent of mechanical tension, but this method does not induce muscle hypertrophy. Therefore, applying BFR during low load exercise may be reliant on mechanical tension in addition to some previously mentioned mechanisms (metabolic accumulation, cell swelling, etc.) to further produce increases in muscle size. Inflating a pressure cuff at the conclusion of one set of high load training would differ from traditional unrestricted high load training in that a greater buildup of metabolites would be pooled within the muscle as a result of the initial exercise bout. If metabolic accumulation can stimulate muscle hypertrophy when maintained after exercise, and independent of mechanical tension, inflating a pressure cuff at the conclusion high load training

may further augment the muscle hypertrophic response seen from high load training itself. However, these alternative mechanisms thought to be responsible for muscle hypertrophy during BFR training may only be important in the absence of sufficient mechanical tension during low load training. During high load training the anabolic stimulus provided from mechanical tension alone is likely enough to maximally stimulate muscle hypertrophy, although this has not been previously studied. By implementing a model that restricts blood flow strictly post exercise, it will allow for the effectiveness of alternative muscle hypertrophic mechanisms to be analyzed without further muscle contraction or mechanical tension. If muscle size is further increased with BFR post exercise, it will demonstrate that these alternative mechanisms can augment muscle growth when maintained after exercise. If there is no augmentation in muscle size, it will demonstrate that mechanical tension may be maximally stimulating muscle growth in itself; or, that metabolic accumulation may be reliant on the presence of mechanical tension.

Purpose

The purpose of the study is to see if restricting blood flow for 3 minutes post exercise can augment muscle hypertrophy independent of additional mechanical tension. Although metabolites will not be directly measured via muscle biopsy, acute data from our laboratory would suggest that prolonged fatigue resulting from the BFR stimulus is evident 3 minutes post exercise, which is likely indicative of metabolic accumulation sustained within the muscle.

Research Question

Will restricting blood flow for 3 minutes post-exercise following high load training produce greater increases in muscle size and strength than high load training itself?

Hypothesis

There will be no difference in muscle size and strength between the conditions performing high load training and those performing high load training followed by BFR. This is hypothesized as it seems likely mechanical tension from high load training is already maximally stimulating the muscle hypertrophic response, in which case adding additional mechanisms may provide no further benefit.

Significance of Study

The implementation of BFR in combination with resistance exercise involves both mechanical tension and alternative mechanisms thought to be involved in muscle hypertrophy (cell swelling, metabolic stress, etc.). However, traditional BFR protocols are not designed to determine the effects of these alternative mechanisms as they are always followed by subsequent bouts of mechanical tension. Therefore, by restricting blood flow at the conclusion of exercise, our design allows us to analyze the muscle hypertrophic effects of these alternative mechanisms without further mechanical tension. High load exercise produces metabolites that may then be pooled within the muscle at the conclusion of exercise with the application of a restriction stimulus. If the pooling of metabolites can further augment muscle growth, it will provide insight that alternative mechanisms may signal muscle hypertrophy when maintained after mechanical tension. If the application of BFR post exercise does not augment muscle growth, it will provide insight that alternative mechanisms occurring during BFR cannot further augment the hypertrophic response from high load training; or, that these alternative mechanisms will not induce muscle growth without the presence of additional mechanical tension.

Assumptions

1. Participants performed as many repetitions as they could.

2. Participants were properly hydrated before testing for muscle thickness.

3. Participants truthfully answered all questions on the health history questionnaire.

Delimitations

1. The results of the study are indicative of the effects on untrained people between the ages of 18-35.

2. The application may be limited to the limbs.

Limitations

 It is possible that a crossover in strength may have occurred where the exercised arm increases the strength of the contralateral arm. However, since both arms were being exercised it is unlikely that this crossover in strength had a large impact on the overall strength of each arm (66).

2. Muscle biopsies were not taken to ensure metabolites were present and elevated during the 3 minute post exercise period; however, acute pilot data from our lab demonstrated a prolonged decrement in torque 3 minutes post exercise which is indicative of an increase in metabolites.

3. We only implemented one set of exercise to avoid having the metabolic stress of blood flow

restriction augment the mechanical tension provided by an additional set of exercise. Despite this, one set of exercise appeared to be sufficient for muscle growth.

4. We retrospectively observed sex differences but were not appropriately powered to analyze them. Future studies could seek to analyze these differences.

5. We cannot infer that trapping metabolites per se is not anabolic given that we were also required to restrict arterial blood flow, potentially limiting the nutrient delivery and associative anabolic signaling(89). Despite this potential limitation, this would seemingly be the only plausible way to pool metabolites post-exercise.

Operational Definitions

1. Blood Flow Restriction (BFR) –Exercise performed with a cuff or wrap placed around the most proximal part of the limb to restrict arterial blood flow and occlude venous return.

2. Arterial Occlusion – The lowest pressure at which no pulse can be detected at the wrist.

3. Muscle Thickness – The distance between the muscle-fat interface and underlying bone will be measured via B-mode ultrasound.

4. 1RM – The maximal load that can be lifted one time with proper form for the dumbbell unilateral elbow flexion exercise.

CHAPTER 2: LITERATURE REVIEW

History of BFR

In 1966 Yoshiaki Sato knelt down at a ceremony and noticed the restricted blood flow in his leg from kneeling produced a numb tingling sensation similar to that felt during resistance exercise (84). He hypothesized a similar restriction of blood flow was playing a role in muscular adaptations seen at the conclusion of weight training. Sato then set out to create a pneumatic device that would allow for the inflation of cuffs to be applied during exercise. Several studies have been published demonstrating the effectiveness of blood flow restriction (BFR) at increasing muscle size during walking (2) and resistance exercise (21). These novel findings have provided the basis for what is today known as BFR training.

Implementation of BFR Stimulus

The pressure applied during BFR should be normalized to the individual and can be estimated based off limb circumference and brachial systolic blood pressure (46). The normalization of BFR can be expressed as a percentage of arterial occlusion and allows for a common stimulus to be applied to all participants. In addition to the applied pressure, the width of the cuff has major implications on the overall restrictive pressure as wider cuffs provide greater restriction stimuli at a given pressure (47). Although a wide range of pressures appear to be effective for the BFR stimulus (49), applying higher pressures may increase the risk of injury, while applying too low of a pressure may be ineffective for stimulating muscle hypertrophy.

Protocols involving BFR training are often classified as either continuous or intermittent. Continuous BFR involves applying the restriction stimulus for the entire duration of exercise, while intermittent allows for unrestricted blood flow during rest periods. There is a distinct difference between these two protocols in that continuous BFR results in a significantly greater metabolic accumulation equivalent to that of high load training; however, intermittent BFR does not (86). This can likely be attributed to the trapping of metabolites within the muscle during continuous BFR, while deflating the cuff intermittently allows metabolites to be flushed out of the muscle. Therefore, continuous BFR results in a greater level of perceived discomfort (18) and makes it a difficult application in conjunction with high load training. To illustrate, Laurentino et al. (44) changed protocols to allow for intermittent BFR as participants were unable to withstand the discomfort of continuous BFR when applied during knee extensor exercises completed with a load corresponding to 80% of their one repetition maximum (1RM).

BFR in the Absence of Exercise

Blood flow restriction applied in the absence of exercise has been shown to attenuate atrophy after periods of disuse (11). The application of BFR in the absence of exercise seems to be effective regardless of the pressure applied as both high (40) and low (39) pressures have successfully attenuated losses in muscular strength. The same protocol used in the previous studies consisting of 5 minute inflations followed by 3 minute deflations, was later shown to have no effect on lactate or EMG activity (48). The authors did note an increase in muscle thickness three minutes after the final deflation which indicated a potential role for cell swelling. Applying BFR without exercise can be a useful tool for people on bed rest or those recovering from major surgery by acting as a stepping stone toward increased physical activity (45).

Aerobic BFR Training

Low intensity aerobic exercise in combination with BFR has been shown to increase muscle size (1, 2, 75, 82) albeit to a lesser extent than when applied in conjunction with low load resistance training. When applied during 2 minute walk intervals lasting 15 minutes in duration, BFR increased quadriceps and lower leg muscle thickness (2, 82). The protocol implemented by Abe et al. was performed at a speed of just 50 m/min and was later shown not to result in increased metabolic accumulation (52). This demonstrates that other mechanisms such as cell swelling and mechanical tension are likely responsible for the hypertrophic effects of slow walking in combination with BFR. Additionally, low intensity cycling performed for 15 minutes at 40% VO_{2 max} increased muscle size, strength, and VO_{2 max} in a BFR group but not control group (1). These studies demonstrate increases in muscle size can be seen across various modalities of aerobic exercise when combined with the BFR stimulus.

Low Load BFR Resistance Training

Blood flow restriction is most commonly incorporated in conjunction with low load training and has been shown to elicit muscle growth in athletic (57), elderly (92), clinical (61) and rehabilitative (71) populations. The standard protocol for BFR training involves 4 sets consisting of 30, 15, 15, and 15 repetitions, respectively, completed with a training load of 20% or 30% 1RM (54). Low load training in combination with BFR has been shown to elicit increases in muscle size comparable to that of high load training (101) and low load training to volitional fatigue (16, 17). These findings support the importance of BFR training in at risk populations as low load training to failure is reliant on training protocols consisting of upwards of 165 repetitions, whereas BFR reduces the required workload to around 75 repetitions (49). To

further support the efficacy of BFR training, perceptual responses tend to be similar to that of low load resistance training to failure (49) and lower (36) or equal (31) to that of high load resistance training.

High Load BFR Resistance Training

High load resistance training in combination with intermittent BFR has been shown to provide no further augmentation to muscle growth when compared to high load unrestricted resistance training (44). In the aforementioned study, participants exercised under complete arterial occlusion and thus intermittent BFR was used due to high levels of discomfort reported by the participants. This is of importance because, as previously mentioned, intermittent BFR results in a significantly lower metabolic accumulation than that of continuous BFR (86). Additionally, exercising with higher loads under complete arterial occlusion probably resulted in a lower total volume of exercise, although this was not reported. A separate study concluded that intermittent BFR, implemented with high load training, resulted in significantly greater increases in lower body strength than high load training without BFR (12). While significant, the group undergoing BFR saw an approximate 3kg greater increase in squat strength which was likely within the error of the measurement as the 2% increase in strength did not exceed the 2.6% coefficient of variation reported with 1RM squat testing (85). The lack of a measure for muscle size makes it further difficult to conclude whether BFR had a significant effect on high load resistance training.

High Load Training with Short Rest Intervals

High load resistance training completed with shorter rest intervals provides a modality of training that although different, provides some similarities to that of BFR training. When very short rest periods are implemented, a greater buildup of metabolites occurs as there is less time for its clearance between sets. To demonstrate, high load training performed with only 10 seconds rest between sets increased lactate levels to greater than 21 mmol/L (38); increases have also been observed at the conclusion of BFR exercise (87). Reducing the rest intervals during resistance training has been shown to produce greater increases in muscle size and strength despite similar volumes of completed work (94), indicating metabolic accumulation may have helped to further augment the increases in muscle size. The impact of metabolic accumulation per se cannot automatically be credited for the increases in muscle size as further mechanical tension was present. It is plausible that a decrease in the amount of time allowed for recovery may have helped to recruit an increased number of type II muscle fibers in the absence of sufficient recovery time.

What Causes Muscle Hypertrophy?

When protein synthesis exceeds protein breakdown a positive net protein balance occurs, that when maintained over time, results in muscle hypertrophy. A variety of physiological adaptations and responses to exercise are thought to be responsible for increases in protein synthesis through the mechanistic target of rapamycin (mTOR) signaling pathway. mTOR exists in 2 complexes appropriately named mTORC1 and mTORC2 and are distinguished by their interaction with the drug rapamycin. mTORC1, which is inhibited by rapamycin, is the major signaling pathway for protein synthesis and can be stimulated by growth factors, amino acids and resistance exercise (59). mTORC2 plays a smaller role in the upward signaling of mTORC1 by allowing for full phosphorylation of protein kinase B (Akt) which is responsible for activating mTORC1 (59). The mTORC1 signaling pathway can be activated through a variety of different mechanisms each taking a unique route based on the initial entry point into the cascade. The mechanisms that are driving muscle hypertrophy are likely all converging on the same mTORC1 pathway and thus may only be necessary for muscle growth in the absence of sufficient mechanical tension; i.e. cell swelling and metabolic accumulation may not be necessary for muscle hypertrophy during high load training due to high levels of mechanical tension already maximizing the hypertrophic response.

The idea that low load training to failure (20-30%1RM), and low load BFR training (20-30%1RM) have both been shown to stimulate muscle hypertrophy provides evidence there are mechanisms other than mechanical tension alone that can promote increases in muscle size (16, 17). One proposed hypothesis is that an accumulation of metabolites, most notably hydrogen ions and lactate, causes a decreased intramuscular pH which may then stimulate muscle growth through a variety of mechanisms (53). However, an increase in metabolites cannot explain the full hypertrophic response occurring during BFR exercise as low intensity walking in combination with BFR elicited muscle growth (2) using a protocol that was later shown not to result in a significant accumulation of metabolites (52). The most robust increases in muscle size are seen when BFR is combined with low load resistance training. When low load resistance exercise is combined with BFR there is an increase in metabolic accumulation, motor unit recruitment and cell swelling. However, it is not presently known whether the robust increases in muscle size from BFR in combination with low load resistance training are a product of increased metabolic accumulation and/or cell swelling per se, or simply increased muscle

12

activation through metabolically fatigued muscle fibers.

Mechanical Tension

Mechanical tension refers to the load placed on a muscle during resistance exercise and is likely the primary stimulator of muscle hypertrophy during high load resistance training. In the absence of heavier loads, mechanical tension alone is not likely sufficient enough to maximally stimulate muscle hypertrophy during low load/low intensity BFR exercise. Although currently unknown, it seems likely that some level of mechanical tension must be present along with another hypertrophic mechanism to produce increases in muscle size from BFR training. Mechanical tension works to stimulate muscle hypertrophy through mechanoreceptors within the muscle that sense levels of tension and respond by activating a protein kinase that eventually activates mTORC1 (59). Mechanical tension during eccentric contractions in rats has been shown to increase phosphorylation of the Tuberous Sclerosis 2 (TSC2) complex, which serves to suppress mTORC1, in which case the phosphorylation of TSC2 would lead to a greater activation of mTORC1 (34). Additionally, greater increases in muscle strength and size are often reported in eccentric as opposed to concentric contractions (93) further supporting the importance of mechanical tension for driving muscle growth. However, when comparing eccentric and concentric isotonic exercise in combination with BFR, greater increases in both muscle size and strength were seen following concentric only training (100). The discrepancy in size and strength was likely due to the increased metabolic accumulation occurring with concentric exercise (73) further suggesting its involvement in muscle growth.

Satellite Cells

It has long been hypothesized that muscle damage may be important for increases in muscle size (102). Mauro (62) accurately predicted satellite cells, located between the

sarcolemma and basement membrane, become activated when stressed, and enter into the sarcoplasm potentially merging with existing myofibers. While BFR exercise may not result in measurable muscle damage (50, 97), proliferation of myogenic stem cells has been shown to occur from BFR training (70). The authors concluded this proliferation of myogenic stem cells may have been due to muscle cell swelling, a hypoxic like stimulus, and/or may have been due to the release of hepatocyte growth factor by means of nitric oxide. The increase in myogenic stem cells is of importance as myonuclei are responsible for supplying mRNA transcripts, and when an insufficient number of myonuceli are present, a muscle cell can no longer grow as the myonucleur domain becomes too large to maintain (91). Work from Stuart Phillips' laboratory recently noted a direct correlation between accretion of satellite cells and muscle growth following high load exercise (5), suggesting they may be responsible to some extent for increases in muscle size. Removing satellite cells, however, has no negative effect on short term muscle hypertrophy (64), demonstrating they are likely only required for long term growth given the myonuclear domain can expand before muscle growth is limited by insufficient myonuclei.

Systemic Hormones

Systemic hormones released during resistive exercise have been long thought to be responsible for increases in muscle size (37) and have been proposed as a mechanism during BFR exercise (58). Work from Stu Phillips' laboratory has recently demonstrated increases in post-exercise systemic hormones likely have little to no effect on muscle size or strength following traditional high load protocols (96). In contrast to traditional high load training, BFR has been shown to increase growth hormone by 290 times that of resting levels (88). This still seems unlikely to have a large impact on muscle hypertrophy as injecting pharmacological doses of growth hormone has been shown to have no positive benefit on increasing muscle size in

adults (6). Further support against systemic hormones playing a large role is the observation that muscle hypertrophy has been shown to occur in BFR but not in non-exercised control limbs (35) despite both limbs being exposed to elevated systemic hormones. Additionally, the most likely hormone to cause anabolic adaptations, testosterone, is either minimally elevated for less than 15 minutes (58) or unchanged (21, 81) post BFR exercise.

Growth Factors

Insulin like growth factor 1 (IGF-1) is secreted by the liver in response to resistance training, and while not mandatory for muscle growth, has been positively correlated (27). An IGF-1 receptor on the sarcolemma is activated by stretching of the muscle and can initiate the mTORC1 cascade as well as the accretion of myonuclei through satellite cell proliferation (24). While systemic IGF-1 does not appear to be elevated substantially (58, 87) or even at all following BFR exercise (20, 21, 77), local IGF-1 produced in skeletal muscle exists in a different isoform (i.e. mechano-growth factor) and may assist in muscle hypertrophy when activated through heavy loading or stretching (83), with the latter possibly occurring during BFR training.

Cell Swelling

Cell swelling occurs when fluid shifts from the plasma into the muscle, but unlike venous pooling of blood, is maintained for at least several minutes post exercise. Blood flow restriction training to failure has shown to increase muscle cell swelling as much as (16) or more so (17) than low load training to failure. The discrepant findings may be due to the load used (30 vs 40% 1RM) whereas heavier loads in conjunction with BFR may promote greater swelling. It has been proposed that the swelling of a muscle can activate a volume sensing G-protein receptor in similar fashion to that of a mechanical sensor during high load training (56). This activation of the G-Protein receptor would then begin the cascade of phosphorylating proteins down the

mTORC1 pathway stimulating muscle growth.

Muscle Activation

The occurrence of muscle hypertrophy is likely dependent upon a high activation of type II fibers which seems to be the one variable constant across all protocols shown to elicit robust muscle growth. While slow walking in combination with BFR has been shown to produce increases in muscle size (2), these increases are only marginal in comparison to muscle growth seen when BFR is combined with low load resistance training (54); which consequently may be attributed to a limited reliance on type II fiber activation. During muscle contraction, type I fibers are preferentially recruited unless a great enough stimulus is needed to recruit both type I and type II fibers (30). Therefore, during low load resistance exercise, type II fibers are rarely recruited which results in a fewer quantity of fibers to be stimulated for growth. Applying BFR has been shown to increase muscle activation to a greater extent than repetition matched low load training during exercise (67, 88) and immediately after rest intervals (99); demonstrating that BFR both increases and prolongs muscular fatigue. When low load training is performed to volitional fatigue, electromyography (EMG) activity is similar (17, 49) or slightly greater (16) than that of BFR training. Interestingly, even with lower EMG activity (55 vs. 65% of maximal isometric strength recorded during the final set), Fahs et al. noted greater increases in lateral thigh muscle size from BFR training. Additionally, when compared to high load training, BFR has been shown to produce equal (86) or slightly lower levels of muscle activation (49, 98). Regardless, the similarity in muscle activation is likely due to an increase in type II fiber activation to assist in lifting a heavier load during high load training; whereas, in BFR training, the prolonged fatiguing of type I fibers requires the additional recruitment of type II fibers to account for the loss of force production (8).

Reduction in Protein Breakdown

An alteration in protein balance can occur by an increase in muscle protein synthesis or a decrease in muscle protein breakdown (MPB). Some negative regulators of muscle growth such as myostatin serve to inhibit mTORC1, while others are directly involved in the degradation of proteins through the ubiquitin proteasome pathway. Basal levels of myostatin messenger RNA (mRNA) expression appear to decline to a slightly greater extent at the conclusion of low load BFR training when compared to high load training completed over 16 training sessions (43). Differential results have been found when assessing MPB after the completion of traditional high load resistance exercise. One study reported elevated MPB at 3 and 24 hours post exercise in a fasted state (79), while another study reported no increase in MPB when assessed 24 hours post exercise in a fasted state (19) despite implementing similar volumes of exercise. Although statistically different outcomes were reported, these values were similar in that Phillips et al reported an 18% increase in MPB while Fry et al. reported a 16% increase. Contrary to traditional high load training, fractional MPB taken in the fasted state at the conclusion of BFR exercise revealed no change at 6 or 24 hours post exercise (26). Since MPB remained unaltered when measured 6 hours post exercise, it would appear any increase in MPB occurring from low load BFR exercise would be limited to shorter post exercise durations than that of high load training.

CHAPTER 3: METHODS

Participants

Sixteen untrained participants were recruited to participate in the study. The sample size was chosen based on an estimated effect size of 0.79 which was averaged from three similar studies [0.53 (32), 0.63 (17) and 1.2 (100)]. Using G*Power software (GPower 3.1), an estimated sample size of 12 people was recommended to appropriately observe statistical significance at the 0.05 alpha level with a power level of 0.8. Inclusion criteria were as follows: (1) must be untrained in the upper body for at least one year; (2) cannot be using tobacco; (3) cannot have had more than one risk factor for thromboembolism (69); (4) and must have been between the ages of 18 and 35. All participants provided informed written consent for this study which was approved by the university's institutional review board.

Study Design

On visit one participants filled out initial paperwork to ensure they were eligible for participation. Following, and in a counterbalanced fashion, participants had one arm assigned as the experimental arm while the other arm served as the control arm. Participants then had their height and body mass measured before undergoing 10 minutes of seated rest. Following rest, participants had their arterial occlusion pressure measured on their experimental arm and were then familiarized with the isokinetic and isometric strength tests. On visit two, participants had their anterior upper arm and thigh (internal control) muscle thickness measured before being tested for maximal isokinetic, isometric, and isotonic (one repetition maximum (1RM)) strength of both arms. Visits 3-26 consisted of exercise training three times for week with each visit separated by at least 48 hours. Visit 27 was held 48-72 hours after the final testing visit and consisted of muscle thickness, and isokinetic, isometric and 1RM strength testing.

Arterial Occlusion

Following 10 min of seated rest, participants were asked to stand and a 5 cm nylon cuff (Hokanson, Bellevue, WA, USA) was placed at the most proximal part of the arm. With an MD6 Doppler probe (Hokanson, Bellevue, WA, USA) at the radial artery, the cuff was inflated by one mmHg increments until a pulse was no longer detected at the wrist. The lowest pressure in which a pulse was no longer present was recorded as the individual's arterial occlusion pressure. The arterial occlusion measure was taken to allow for the restrictive stimulus to be made relative to each individual as suggested previously (47).

Isometric and Isokinetic Strength

Participants were seated on a dynamometer (Biodex Medical Systems, Shirley, New York, USA) with the seat and lever arm adjusted appropriately and the settings recorded and standardized for all future tests. The dynamometer was adjusted for each individual and all settings were recorded to ensure a similar testing protocol throughout. After weighing the individuals arm to correct for gravity, participants performed 3 successive isokinetic contractions at 180°/s and then rested for 90 seconds before repeating the test again at the same speed. After another 90 seconds of rest participants performed the same procedure involving 2 sets of 3 isokinetic contractions at 60°/s. Following another 90 seconds of rest, participants performed isometric testing in the same position. The lever arm was locked into place at 60° and participants performed 2 maximal isometric contractions each lasting 3 seconds in duration and separated by 1 minute of rest. The highest value for each test was recorded as the maximum peak torque.

One-Repetition Maximum (1RM) Strength

After a brief warmup consisting of 7-10 repetitions with approximately 30% of the individuals estimated 1RM, the load was increased to approximately 70% 1RM and one repetition was performed. After increasing the load to an estimated 90% 1RM individuals performed a 1RM attempt. The load was then progressively increased until the individual could no longer perform the exercise through a full range of motion with proper form. All 1RM attempts were separated by approximately 90 seconds rest and were performed with the individuals back and heels against a wall to ensure strict form. All 1RMs were measured to the nearest 0.5 kg and were usually obtained in around 5 attempts. The 1RM tests were performed pre and post exercise, as well as during the 13th (i.e. the midpoint) visit in order to readjust the training load.

Muscle Thickness

An Aloka SSD-500 B-mode ultrasound (Aloka Co. Ltd., Tokyo, Japan) was used to measure the distance between the muscle-fat and muscle-bone interface by an experienced tester. All images were printed and analyzed by the same person who was blinded to the condition. Three images were taken at each of three sites including 50%, 60%, and 70% the distance between the lateral epicondyle and the acromion process. An additional measure of thigh muscle thickness was taken at 50% of the distance between the lateral epicondyle of the femur and the greater trochanter and was used to assess the stability of the measurement over time. Participants were asked to refrain from any planned exercise within 24 h of muscle thickness measures. The minimal difference (i.e. reliability) needed to be considered real for the anterior portion of the upper and lower arm was calculated at 0.2 cm prior to the investigation using the procedure detailed previously by Weir (95).

Arm Circumference

The distance from the acromion process to the lateral epicondyle was measured with a standard tape measure and a mark was made 10 cm proximal to the lateral epicondyle. Circumference measures were taken on both arms every training visit prior to exercise.

Training Protocol

In a counterbalanced fashion individuals were assigned one arm to serve as the experimental arm and one arm to serve as the control arm. On training visits individuals performed one set of standing elbow flexion exercise to volitional fatigue using a load corresponding to 70% of the predetermined 1RM for that arm. All exercise was performed to the beat of a metronome allowing 1 second for the concentric and 1 second for the eccentric portion of the exercise. During each training session, individuals alternated which arm exercised first and 5 minutes of rest preceded exercise of the contralateral arm. Both arms exercised with a 5 cm nylon cuff (Hokanson, Bellevue, WA, USA) placed at the most proximal part of the arm. Upon completion of the final repetition, the control arm had the pressure cuff removed immediately, while the experimental arm had the cuff inflated to 70% of their predetermined arterial occlusion pressure for 3 minutes. During the 3 minute post-exercise period individuals were required to remain standing with their arms kept loosely at their side.

Ratings of Perceived Discomfort

The Borg (CR10+) scale was used to assess ratings of discomfort before, immediately post, and 1,2, and 3 min post-exercise. The scale was explained in depth to all participants on the initial training visit and all participants fully understood the scale. As described previously (51), participants were asked "What are your worst experiences of discomfort? 'Maximum discomfort (rating of 10)' is your main point of reference; it is anchored by your previously experienced

worst discomfort. The worst discomfort that you have ever experienced, the 'Maximum discomfort' may not be the highest possible level of discomfort. There may be a level of discomfort that is still stronger than your 10; if this is the case, you will say 11 or 12. If the discomfort is much stronger, for example, 1.5 times 'Maximum Discomfort' you will say 15; any questions?"

Statistical Analysis

Using the SPSS 20 statistical software package (SPSS Inc., Chicago, IL) a 2 (condition) x 3 (time) repeated measures analysis of variance (ANOVA) was used to determine significant changes in 1RM strength. If an interaction was present a one ANOVA way was used to compare differences across time for each condition and a paired t test was used to compare differences between conditions at each time point. If no interaction was present main effects of time and condition were interpreted. Additionally, a 2 (condition) x 2 (time) repeated measures ANOVA was used to compare differences for muscle thickness, isometric strength, isokinetic strength, volume, repetitions, and circumference. If there was an interaction paired t tests were used to compare differences across time and to compare differences between conditions at each time point. If no interaction was present main effects of time and to compare differences between conditions at each time point. If no interaction was present main effects of time and compare differences across time and to compare differences between conditions at each time point. If no interaction was present main effects of time and condition were interpreted. Finally, a Wilcoxin signed-rank test was used to compare ratings of discomfort between conditions at each of the 5 time points. The level of significance will be set at $p \leq 0.05$ for all statistical tests.

CHAPTER 4: RESULTS AND DISCUSSION

All data are reported as means (standard deviations) with the exception of discomfort which is reported as 50th percentile (25th percentile, 75th percentile).

Demographics

A total of 16 individuals were recruited for participation in the study. Three individuals withdrew for personal reasons unrelated to participation in the study, and thus data for 13 individuals (6 males and 7 females) was included in the analysis (Table 1). The average age, height and body mass, respectively, were 22 (3) years, 169.1 (9.4) cm and 76.2 (20.0) kg (Table 2). The average arterial occlusion pressure measured before exercise was 152 (25) mmHg which corresponded to 106 (17) mmHg applied as the 70% arterial occlusion post-exercise.

		Age	Height	Body Mass	Total AOC	70% AOC
ID	Sex	(years)	(cm)	(kg)	(mmHg)	(mmHg)
2	Μ	20	161.6	56.6	111	78
3	F	20	165.7	48.1	140	98
4	F	23	161.7	53	143	100
5	F	21	163.8	95.4	154	108
7	Μ	26	183.7	104.2	164	115
8	F	21	168.4	84.2	205	144
9	F	21	153.8	72.3	180	126
10	F	23	162.6	62	133	93
12	Μ	21	168.2	89	134	94
13	F	24	177.7	66.2	138	97
14	F	25	166.8	73.4	163	114
15	F	20	183.6	113.7	182	127
16	F	32	180.8	73.7	136	95

Table 1. Individual Demographics

AOC = arterial occlusion pressure, M=male, F=female

Muscle Thickness

There was no condition x time interaction (p=0.31), main effect of condition (p=0.71) or main effect of time (p=0.19) at the 50% site of the anterior upper arm (Table 3, Figure 1). At the 60% site, there was a condition x time interaction (p=0.03) with the control condition increasing from pre to post (p=0.042), however, no change was observed in the experimental condition from pre to post (Table 3, Figure 1, p=0.74). Additionally, when examining post muscle thickness at the 60% site, there was a trend toward greater muscle thickness in the control vs. experimental condition (p=0.06). At the 70% site, there was no condition x time interaction (p=0.90) or main effect of condition (p=0.177), however there was a main effect of time (Table 3, Figure 1, p=0.006) with muscle thickness increasing from pre to post. For thigh muscle thickness which served as the internal control [control pre: 3.9 (1.4) cm, control post: 3.9 (1.3) cm; experimental pre: 4.0 (1.2) experimental post: 4.1 (1.4)], there was no condition x time interaction (p=0.25), main effect of condition (p=0.55) or main effect of time (p=0.29) suggesting our measurements were stable across time. Interpreting these changes within the context of our reliability, we are confident that we have indeed measured muscle growth as opposed to edema (14) because we did not see increases in arm circumference, did not observe any pre exercise discomfort during any of the training sessions, and performed the post-training muscle thickness measurement 48-72 hours after training which has previously demonstrated to be ample time for swelling to subside when assessed at the fiber level (70).

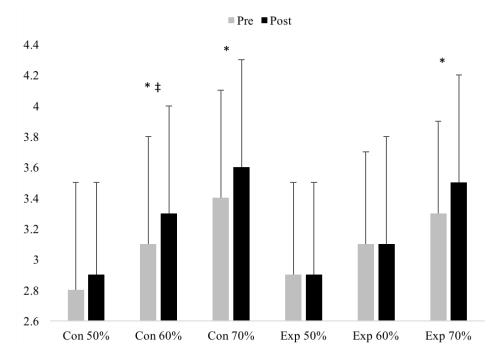
Table 2. Muscle Thickness

Control			Experimental				
Location	Pre	Post	Δ	Pre	Post	Δ	
50%	2.8 (0.7)	2.9 (0.6)	0.1	2.9 (0.6)	2.9 (0.6)	0.0	
60%	3.1 (0.7)	3.3 (0.7)*	0.2	3.1 (0.6)	3.1 (0.7)	0.0	
70%	3.4 (0.7)	3.6 (0.7)*	0.2	3.3 (0.6)	3.5 (0.7)*	0.2	
Thigh	3.9 (1.4)	3.9 (1.3)	0.0	4.0 (1.2)	4.1 (1.4)	0.1	

All values (cm) are presented as mean (standard deviation) *statistically significant from pre value

When analyzing individual responses at or exceeding 0.2 cm (error of measurement), there were a greater number of participants displaying meaningful increases in the control condition at each the [50% (control = 46% vs. experimental = 23%), 60% (control = 53% vs. experimental = 15%), and 70% sites (control = 61% vs. experimental = 38%) Table 4]. Individual responses displaying the within participant difference in muscle growth between the control and experimental condition at each site are displayed in Figure 2. That is, for each of the 3 sites (i.e. 50, 60, 70%) on all 13 individuals (total of 36 calculations), the pre to post change in muscle thickness of the control condition was subtracted from that of the experimental condition (Δ experimental - Δ control). Therefore, a negative value demonstrates a more advantageous effect of muscle size for the control condition, while a positive value demonstrates a similar response between the experimental and control conditions. Furthermore, the individual pre to post changes for the experimental and control conditions are displayed in Figure 3.

Figure 1. Muscle Thickness



Values are expressed as mean (standard deviation). The 50%, 60% and 70% sites indicate the location of muscle thickness measured as the distance from the lateral epicondyle to the olecranon process. con = control, exp = experimental, *significantly different from pre, \ddagger condition x time interaction with the control condition trending toward being greater than the experimental at the post measure (p=0.06).

	Control	Experimental
50%	6	3
60%	7	2
70%	8	5

Table 3. Meaningful Increases in Muscle Thickness

Values are individual participants meeting or exceeding the error (0.2 cm) of the measurement. The 50%, 60% and 70% sites indicate the location of muscle thickness measured as the distance from the lateral epicondyle to the olecranon process.

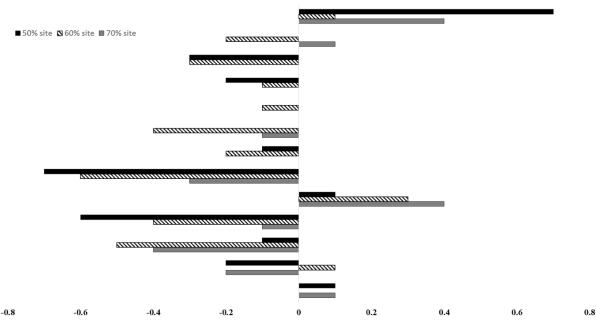


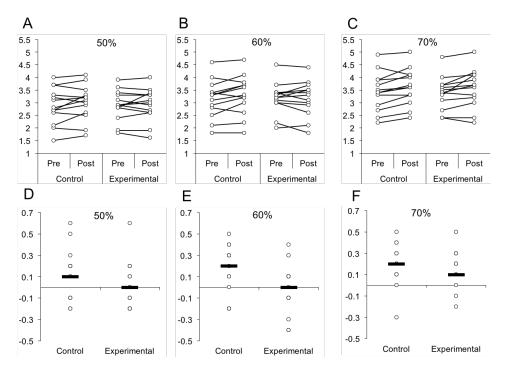
Figure 2. Within Subject Differences in Muscle Thickness Between Conditions



Values are calculated for each individual at each location using the equation (Δ experimental - Δ control). A positive value favors muscle growth in the experimental condition, a negative value favors muscle growth in the control condition, and a value of 0 indicates a similar response between the control and experimental condition.

The 50% site was the only measured site where muscle growth was not observed at the group level in either the control or experimental condition. It has previously been suggested that the application of a pneumatic cuff during resistive exercise may attenuate growth of muscle tissue placed under the cuff (15, 35). While, we applied a cuff that was not inflated, there still was some restriction placed on the anterior upper arm, particularly during the concentric portion of the exercise. Even so, muscle growth has been shown to occur in exercised tissue located under restriction (42), leaving open the possibility that the volume of exercise performed in the present study (1 set) was not sufficient to activate the more proximally located portion of the elbow flexors. We also cannot rule out the possibility that the 50% location was not activated enough during the elbow flexion exercise given the proximal location that was measured.

Figure 3. Individual Changes in Muscle Thickness



A-C illustrate pre to post values (cm) at each the 50, 60 and 70% sites measured for muscle thickness. D-F illustrate the pre to post changes in muscle thickness (cm) with circles representing each individual and the solid black line representing the median pre to post change (some circles may represent more than one individual, if they both had similar median differences); exp=experimental, con=control.

The 60% location clearly displays that the experimental condition appeared to be attenuated by the post-exercise application of BFR (Figure 1, Table 3). While statistically significant at the group level, this was also observed at each of the 3 measured sites when examining individual responses (Figure 2, Figure 3). While we hypothesized there would be no difference in muscle growth between conditions, we thought it would be plausible that the experimental arm would see a greater increase in muscle size given the trapping of metabolites post-exercise. For example, lactate has been shown to induce hypertrophy when administered *in vitro* and in mouse models performing treadmill exercise (72). While unexpected, the attenuated growth in the experimental condition may be partially explained by the location and quantity of

reactive oxygen species produced. For example, reperfusion results in a drastic increase in reactive oxygen species produced within the mitochondria which may serve to inhibit upstream activators of the mechanistic target of rapamycin (mTOR), whereas traditional exercise produces reactive oxygen species at locations along the sarcolemma (e.g. NADPH and Xanthine oxidases) (60) thought to be involved in mechanotransduction (33). Additionally, it has been shown that muscle contraction during BFR alleviates some of the oxidative stress caused by vascular occlusion (22, 23), and therefore, it is possible that the magnitude of oxidative stress caused by high load training combined with post-exercise occlusion exceeded that which has been speculated to be beneficial (7, 60, 65).

In addition to the possibility that metabolites may have been detrimental through oxidative stress, it is possible that metabolic accumulation may have been detrimental through activation of the energy sensing AMP-activated protein kinase (AMPK) complex, which serves to decrease protein synthesis and elevate proteolysis (28). We cannot strictly attribute the detrimental effects observed to the trapping of metabolites, however, as we also restricted arterial blood flow for a 3 minute post-exercise period. It has previously been suggested that an increase in post-occlusive blood flow may be responsible for some of the adaptation provided by blood flow restriction (76), and while this has been refuted elsewhere (25), the elevation in blood flow caused by sodium nitroprusside did not match the immediate post-exercise elevation in blood flow caused by BFR. This restriction of blood flow could have potentially limited post-exercise nutrient delivery and associative anabolic signaling (89). The limited arterial blood flow could also have induced a state of hypoxia which could serve to increase protein degradation through up-regulation of the E3 ligases muscle ring finger-1 (MURF1) and muscle atrophy F box (MAFbx) associated with the ubiquitin proteasome system (10). Despite this finding, the increase

in E3 ligases may be attributed to remodeling associated with increased protein synthesis (4) occurring from the synergistic ablation procedure as opposed to the hypoxic environment. While it has previously been hypothesized that BFR may augment muscle growth by decreasing myostatin mRNA (43), it is possible that our protocol may have actually increased myostatin mRNA from the hypoxic stimulus (29), resulting in a decrease in protein synthesis through inhibition of protein kinase B (Akt), an upstream activator of mTOR. The increase in myostatin mRNA resulting from hypoxia appeared to diminish, however, with the presence of functional overload (10). Additionally, the present study differs from systemic hypoxia in that we analyzed a localized hypoxic-like stimulus specific to the arm, which may have produced different results.

It could be hypothesized that a larger muscle would be capable of producing a greater number of metabolites, and therefore the larger amount of muscle mass present in males may produce different results from the post-exercise application of BFR. For this reason, we retrospectively analyzed differences between males and females in relation to changes in muscle thickness (Figure 4, Table 5), but did not perform additional statistical analyses because we were not powered to do so. However, when examining Figure 4, there were only two experimental conditions that decreased from pre to post in males, whereas 10 conditions decreased in females. Additionally, when analyzing Table 5, it would appear that females were more negatively impacted in the experimental condition when compared to males.

This detrimental effect of post-exercise BFR being specific to females is difficult to explain, but may be related to differences in metabolites and/or fiber type composition between genders. For example, given that females possess a larger percentage of type I fibers in comparison to males, it is possible that females produced a greater number of mitochondrial reactive oxygen species (80). Despite the production of endogenous antioxidants within the

mitochondria(80), it is possible that the ratio of reactive oxygen species exceeded that of antioxidants, thus resulting in detrimental levels of oxidative stress (7, 60, 65), particularly when produced within the mitochondria (60). Another possible explanation is that because males express more type II muscle fibers they produce a greater amount of lactate during exercise. If lactate is indeed anabolic as previously suggested (72), it is possible that the anabolic effects of lactate were sufficient to overcome the catabolic effects caused by restricting blood flow for 3 minutes post-exercise.

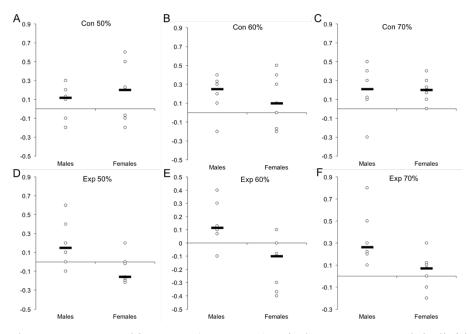


Figure 4. Sex Differences in Muscle Thickness

Values are expressed in cm as (post – pre). Circles represent each individual change and the solid black line represents the median pre to post change (some circles may represent more than one individual, if they both had similar median differences); exp = experimental, con=control.

	Pre 50%	Post 50%	Δ
Males Control	3.4 (0.4)	3.5 (0.4)	0.1
Male Experimental	3.2 (0.4)	3.3 (0.3)	0.1
Females Control	2.4 (0.5)	2.5 (0.5)	0.1
Females Experimental	2.6 (0.5)	2.5 (0.5)	-0.1
	Pre 60%	Post 60%	Δ
Males Control	3.6 (0.5)	3.8 (0.4)	0.2
Male Experimental	3.5 (0.5)	3.7 (0.3)	0.2
Females Control	2.6 (0.5)	2.8 (0.6)	0.2
Females Experimental	2.8 (0.5)	2.7 (0.5)	-0.1
	Pre 70%	Post 70%	Δ
Males Control	4.0 (0.5)	4.2 (0.4)	0.2
Male Experimental	3.8 (0.5)	4.1 (0.4)	0.3
Females Control	2.9 (0.5)	3.1 (0.4)	0.2
Females Experimental	3.0 (0.5)	3.0 (0.5)	0

Table 4. Sex Differences in Muscle Thickness

All values (cm) are expressed as mean (standard deviation).

One-Repetition Maximum Strength (1RM)

There was no condition x time interaction (p=0.94) or main effect of condition (p=0.77),

however, there was a main effect of time (p<0.001) with 1RM strength increasing from pre to

mid (P<0.001), mid to post (Table 6, p=0.002), and pre to post (p<0.001).

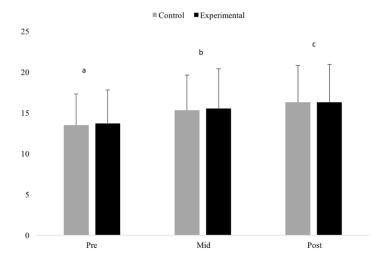


Figure 5. One Repetition Maximum (1RM) Strength

Values (kg) are presented as mean (standard deviation). There were no significant differences between conditions. Letters indicate significant differences from one another.

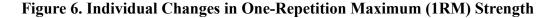
The increase in 1RM strength was similar for both the control and experimental condition, which is not surprising given that both arms performed the identical protocol throughout the study duration. The dissociation between muscle size and strength has been documented previously (66), and is likely related to the principle of specificity. Whereas muscle growth is largely reliant on fatiguing the muscle (66) and increasing muscle activation (68), increases in 1RM strength can be accomplished by adhering to the principle of specificity and performing resistance exercise at or near an individuals 1RM for that particular exercise (74). The individual responses in 1RM strength appear to be fairly consistent across the entire study population (Figure 6). These results demonstrate that pooling metabolites post-exercise did not appear to have any effect at the individual level either.

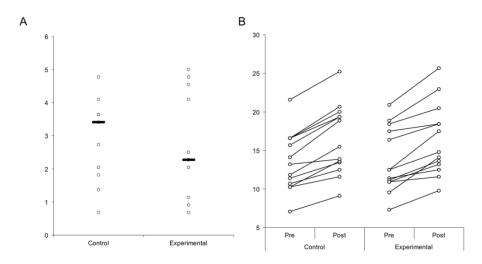
	Pre ^a	Mid ^b	Post ^c
Control	13.5 (3.8)	15.3 (4.3)	16.3 (4.5)
Experimental	13.7 (4.1)	15.5 (4.9)	16.3 (4.6)

Table 5. One Repetition Maximum (1RM) strength

Values are expressed in kg as mean (standard deviation). Letters indicate significant differences. Both conditions increased from pre to mid and mid to post with no differences between conditions.

The differences in 1RM strength were quite large with some individuals increasing 1RM strength by less than 1 kg, while others increased by more than 5 kg (Figure 5B). This variability in individual responses has been observed previously in a large cohort of individuals also performing elbow flexion exercise (32). Furthermore, males in the study saw greater absolute increases in 1RM strength (3.4 vs. 1.6 kg); however, when expressed relative to pre-training values males and females saw similar relative increases (19 vs. 15%) (Table 7), which is supportive of previous research (32).

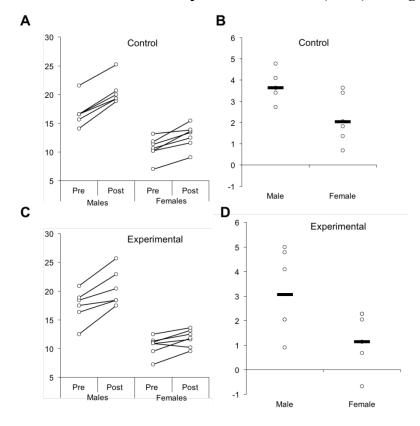




Data are expressed in kg. A. Individual changes in 1RM strength. B. Pre to post changes in 1RM strength. Circles represent individual changes and the solid line represents the group median.

The differences in 1RM strength were quite large with some individuals increasing 1RM strength by less than 1 kg, while others increased by more than 5 kg (Figure 5B). This variability in individual responses has been observed previously in a large cohort of individuals also performing elbow flexion exercise (32). Furthermore, males in the study saw greater absolute increases in 1RM strength (3.4 vs. 1.6 kg); however, when expressed relative to pre-training values males and females saw similar relative increases (19 vs. 15%) (Table 7). This is supportive of previous research examining sex differences in elbow flexion 1RM strength (32).

Figure 7. Sex Differences in One-Repetition Maximum (1RM) Strength



All values are expressed in kg. Figures A and C illustrate pre to post changes for each specific individual. Figures B and D illustrate changes in 1RM strength (post – pre) with circles illustrating individual responses and the solid line depicting the median value (some circles may represent more than one individual, if they both had similar median differences).

Given the differences in the control and experimental conditions present in females for

muscle thickness, additional individual plots were created to compare pre to post differences in 1RM strength between conditions and across sex (Figure 7). We did not observe any clear pattern for 1RM strength between conditions for either males or females. Specifically, the detrimental effect observed in the experimental condition of females did not result in reductions in 1RM strength. While Table 7 illustrates a trend toward the experimental condition resulting in greater increases in 1RM strength among both males and females, the individual plots reveal that this may be largely driven by the responses of one male and one female included in the analysis (Figure 7).

Table 6. Sex Differences in One-Repetition Maximum (1RM) Strength

	Pre	Post	Δ
 Male Control	16.8 (2.5)	20.5 (2.3)	3.7 (22%)
Male Experimental	17.4 (2.8)	20.5 (3.1)	3.1 (17%)
Female Control	10.6 (1.8)	12.7 (2.0)	2.1 (19%)
 Female Experimental	10.5 (1.6)	11.7 (1.4)	1.2 (11%)

Values (kg) are expressed as mean (standard deviation).

Isometric and Isokinetic Strength

For isokinetic strength at 180° /s there was no condition x time interaction (p=0.40) nor was there a main effect of condition (p=0.253) or time (p=0.975) (Table 8, Figure 8). For isokinetic strength at 60°/s there was no condition x time interaction (p=0.81) nor was there a main effect of condition (p=0.138) or time (p=0.562) (Table 8, Figure 8). With regard to isometric strength, there was no condition x time interaction (p=0.285) nor was there a main effect of condition (p=0.507) or time (p=0.963) (Table 8, Figure 8).

 Table 7. Isometric and Isokinetic Strength

	Control		Experimental	
	Pre	Post	Pre	Post
Isokinetic 180°/sec	36.0 (10.6)	35.7 (11.7)	37.0 (10.3)	37.3 (11.2)
Isokinetic 60°/sec	42.1 (14.3)	42.3 (14.4)	43.8 (13.5)	44.3 (14.2)
Isometric at 60°			()	49.7 (16.1)

All values (Nm) are expressed as mean (standard deviation)

There were no differences observed from pre to post for either the control or experimental condition for isometric or isokinetic strength. The lack of improvement in isometric or isokinetic strength demonstrated in the present study may be largely attributed to the principle of specificity. Given that individuals trained with isotonic exercise it would be expected that isotonic (1RM) strength would increase to a greater extent than that of isometric or isokinetic strength. One set to volitional fatigue has previously been demonstrated to increase isometric strength of the knee extensors to the same extent as three sets (66), however, this may be different in the elbow flexors. To illustrate, a previous study found increases in isometric strength of the elbow flexors with 3 sets (32), suggesting that more repetitions may be necessary to increase isometric/isokinetic strength of the elbow flexors. This hypothesis is speculative, however, as no previous studies to our knowledge have compared isometric/isokinetic strength of the elbow flexors.

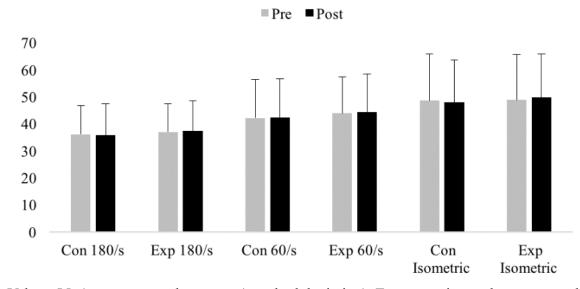


Figure 8. Isometric and Isokinetic Strength

Values (Nm) are expressed as mean (standard deviation). Exp=experimental, con=control.

When examining individual changes in torque (Figure 9) there did not appear to be any trend for the control or experimental condition for any of the tests. As depicted at both the group and individual level, there was an increase in torque production as the speed of the contraction reduced, ultimately resulting in the greatest torque production during the isometric test. The force velocity relationship illustrates that at higher velocities there is less time to apply torque, thus explaining why the lowest torque production was observed during the fastest isokinetic test.

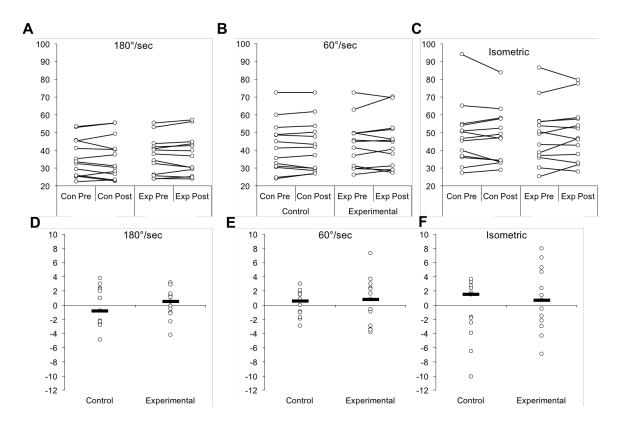


Figure 9. Individual Changes in Isometric and Isokinetic Strength

All values are expressed in nm. A, B and C illustrate individual pre to post changes. Figures D, E and F are presented as (post – pre) with circles representing individual changes and solid lines representing the group median.

Volume

There was no condition x time interaction for volume (p=0.94) nor were there main

effects of condition (p=0.74) or time (p=0.88). The total volume completed by the control and

experimental arms did not differ during the first 12 or last 12 sessions, nor were there differences

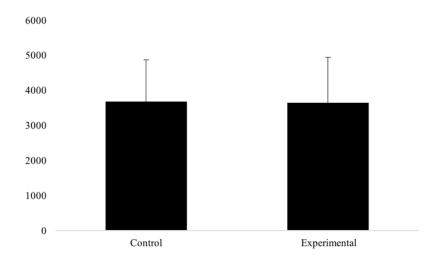
in total exercise volume (Table 9, Figure 10).

First 12 Sessions	Last 12 Sessions	Total
1834.3 (577.9)	1843.9 (642.2)	3678.3 (1183.9)
1812.6 (584.9)	1826.0 (740.1)	3638.7 (1297.8)
	1834.3 (577.9) 1812.6 (584.9)	1834.3 (577.9) 1843.9 (642.2)

Table 8. Exercise Volume (repetitions x load)

All values are in kg and are expressed as mean (standard deviation).

Figure 10. Exercise Volume



Values (kg) are expressed as means and standard deviations.

The attenuation of growth that was present in the experimental condition occurred despite performing the same volume of exercise and relative load as the control condition. It has previously been demonstrated that more volume does not always result in greater muscle growth (63) as the benefits gained from a resistance training protocol are undoubtedly finite. With the exclusion of studies implementing BFR, no previous study has demonstrated differential muscle growth involving two protocols implementing the identical protocol (i.e. volume, sets, relative load. Previous studies matching work and volume through the use of different relative loads and repetitions have demonstrated differences in muscle protein synthesis (41) demonstrating that volume is not necessarily the most important determinant for the hypertrophic potential of an exercise protocol. Nonetheless, in addition to exercise volume, the present study employed the identical exercise protocol involving one set performed to volitional fatigue.

Repetitions

There was no condition x time interaction for repetitions (p=0.74) or main effect of condition (p=0.78), however there was a main effect of time (p=0.01) with repetitions completed

decreasing from the first 12 sessions to the final 12 sessions (Table 10, Figure 11).

Table 9. Repetitions

	First 12 Sessions	Last 12 Sessions*	Total
Control	195 (41)	171 (30)	366 (65)
Experimental	194 (52)	167 (45)	362 (89)
Values are in kg and a	re expressed as mean (standard deviation) *stati	stically different from the

first 12 sessions

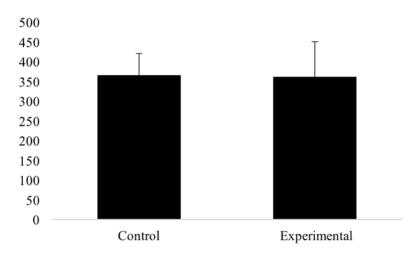


Figure 11. Total Repetitions

Values are expressed as means and standard deviations.

Unsurprisingly, both the control and experimental conditions performed nearly the identical number of repetitions over the course of the study (Table 10, Figure 11). When breaking down the number of repetitions comparing the first 12 visits against the last 12 visits, there were more repetitions completed in the last 12 sessions (Table 10). This can be explained by the retesting of 1RMs that were performed prior to training on visit 13. The increase in the load resulted in a decrease in the number of repetitions necessary to reach volitional fatigue. Given that the number of repetitions, relative load, volume, and fatiguing sets of exercise were all similar, the difference in muscle growth observed must be attributed to the application of post-exercise BFR.

Arm Circumference

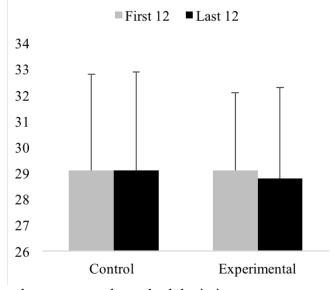
There was no condition x time interaction (p=0.27) nor was there a main effects of condition (p=0.47) or time (p=0.27) for arm circumference (Table 11, Figure 12).

Table 10. Arm Circumference

		First 12 Sessions	Last 12 Sessions	Total Average
	Control	29.1 (3.7)	29.1 (3.8)	30.2 (3.7)
	Experimental	28.8 (3.0)	291 (3.5)	30.0 (3.3)
T 7 1	• •	1	(, 1 1 1 · · · ·)	

Values are in cm and are expressed as mean (standard deviation)

Figure 12. Arm Circumference



Values (cm) are expressed as means and standard deviations

The assessment of arm circumference was primarily used to test whether the muscle growth that was present was not due to edema. Since circumference stayed constant from the first 12 sessions to the last 12 sessions this would suggest that our measure of muscle size was not largely impacted by edema.

Discomfort

The median values over the first 12 and final 12 training sessions were calculated for each individual at all 5 time points (pre, post, 1, 2, 3 min). Each individual's median values for the first 12 and last 12 sessions were then used to calculate the median discomfort at the group level. During the first 12 sessions, there were no differences in the median discomfort at pre (p=0.99) or immediately post-exercise (p=0.28), however the median discomfort was significantly greater in the experimental arm at 1 (p=0.002), 2 (p=0.002) and 3 (p=0.001) min

post-exercise (Table 12, Figure 13). The same results were observed for median values during the final 12 sessions of the study with no differences observed at pre (0.31) or immediately post-exercise (p=0.52), however the median discomfort was significantly greater in the experimental arm at 1 (p=0.003), 2 (p=0.002) and 3 (p=0.002) min post-exercise (Table 12, Figure 13).

Table 11. Borg (CR10+) Ratings of Discomfort

	First 12 sessions		Last 12 sessions	
	Control Experimental		Control	Experimental
Pre	0 (0,0)	0 (0,0)	0 (0,0)	0 (0,0)
Post	0.5 (0.3, 0.6)	0.5 (0.3, 0.7)	0.5 (0.3, 1.0)	0.5 (0.3, 1.5)
1-min post	0.5 (0.4, 0.8)	1.2 (0.8, 2.0) *	0.5 (0.3, 1.0)	1.0 (0.7, 2.5) *
2-min post	0.5 (0.1, 0.8)	2.0 (1.2, 3.2) *	0.5 (0.1, 1.0)	2.0 (0.9, 3.0) *
3-min post	0.3 (0.0, 0.7)	2.5 (1.5, 3.5) *	0.3 (0.1, 1.0)	2.5 (1.2, 3.0) *

All values are presented as median (25th percentile, 75th percentile) for all individuals across the first 12 and last 12 training sessions. *significantly different than control value at same time point

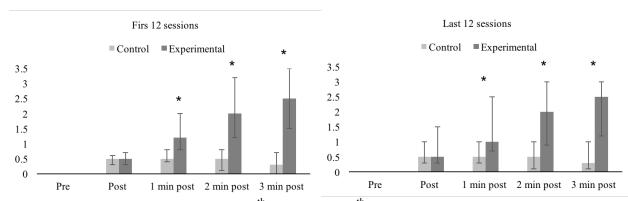


Figure 13. Borg (CR10+) Ratings of Discomfort

All values are presented as median (25th percentile, 75th percentile) for all individuals across the first 12 and last 12 training sessions. *significantly different than control value at same time point

The levels of discomfort following 3 minutes of post-exercise BFR in the current study (2.5) were similar to what has been observed using a higher restrictive pressure in the upper body (discomfort ranged from 3 to 4) (13). This similarity would be expected given both studies incorporated elbow flexion exercise using similar restrictive pressures (70% vs. 90% arterial occlusion). Additionally, Loenneke et al. observed similar ratings of discomfort when applying BFR in the absence of exercise. This was observed following 3 continuous minutes of BFR (discomfort = 2.5) (51) and following cycles of 5 minute inflations and 3 minute deflations (discomfort = 2.7) (48). Anecdotal reports from participants performing previous studies in our laboratory would suggest that discomfort is greater as the period of BFR is prolonged, and this discomfort is alleviated to some extent by further muscle contraction.

CHAPTER 5: CONCLUSION

The main purpose of the study was to see if performing one set of high load resistance exercise could be augmented by applying blood flow restriction (BFR) for 3 minutes post-exercise. Measures of isotonic (1RM), isometric and isokinetic strength were also analyzed.

Hypotheses

1. There will be no difference in muscle size and strength between the control and experimental conditions.

This hypothesis did not appear to be supported by the data. While there were no significant differences between the experimental and control conditions at either the 50% or 70% sites, there was an interaction at the 60% site demonstrating muscle growth to be attenuated in the experimental condition. Additionally, when analyzing within subject responses between conditions, there appeared to be an attenuation of muscle growth across all sites in the experimental arm in comparison to the control conditions. Furthermore, retrospective sex comparisons demonstrated that, in comparison to the control condition, applying BFR post high load exercise appeared to be more detrimental in females when compared to males.

2. There will be no difference in muscle strength between the control and experimental conditions

This hypothesis was supported by the data given there were no differences in the control or experimental condition at any of the time points examined. Both conditions increased similarly from pre to mid and from mid to post

Significance

The application of BFR allows for individuals to increase muscle size through the use of low load protocols that would otherwise not result in muscle growth. While BFR is thought to work through the pooling of metabolites, previous studies have not been designed to tease out the importance of metabolic accumulation on muscle hypertrophy. All previous BFR studies have been used exclusively with multi-set resistance training protocols, thus allowing for metabolites to augment muscle activation of subsequent sets. By applying BFR at the conclusion of one set of high load training, this study may provide some insight that: 1) metabolites may not have anabolic properties *per se*, and may actually be detrimental for muscle growth when prolonged at the conclusion of high load exercise; 2) the immediate increase in blood flow occurring at the conclusion of exercise may be of great importance for inducing muscle growth; and/or 3) metabolites have anabolic properties but this was masked by the restriction of blood flow post-exercise.

Future Research

Future studies could seek to determine the differences in intramuscular metabolites caused by restricting blood flow for 3 minutes post-exercise. Additionally, future studies could

seek to determine why the application of post-exercise BFR attenuated muscle growth, and further, why this appeared to occur predominantly in females. Finally, studies may seek to determine if the detrimental effects of applying BFR post high-load exercise are still evident when applied post low-load exercise.

BIBLIOGRAPHY

- 1. Abe T, Fujita S, Nakajima T, et al. Effects of Low-Intensity Cycle Training with Restricted Leg Blood Flow on Thigh Muscle Volume and VO2MAX in Young Men. *J Sports Sci Med* 2010;9(3):452–8.
- 2. Abe T, Kearns CF, Sato Y. Muscle size and strength are increased following walk training with restricted venous blood flow from the leg muscle, Kaatsu-walk training. *J Appl Physiol Bethesda Md 1985* 2006;100(5):1460–6.
- 3. American College of Sports Medicine. American College of Sports Medicine position stand. Progression models in resistance training for healthy adults. *Med Sci Sports Exerc* 2009;41(3):687–708.
- 4. Baehr LM, Tunzi M, Bodine SC. Muscle hypertrophy is associated with increases in proteasome activity that is independent of MuRF1 and MAFbx expression. *Striated Muscle Physiol* 2014;5:69.
- 5. Bellamy LM, Joanisse S, Grubb A, et al. The acute satellite cell response and skeletal muscle hypertrophy following resistance training. *PloS One* 2014;9(10):e109739.
- 6. Birzniece V, Nelson AE, Ho KKY. Growth hormone and physical performance. *Trends Endocrinol Metab TEM* 2011;22(5):171–8.
- Bloodsworth A, O'Donnell VB, Freeman BA. Nitric oxide regulation of free radical- and enzyme-mediated lipid and lipoprotein oxidation. *Arterioscler Thromb Vasc Biol* 2000;20(7):1707–15.
- Burd NA, Mitchell CJ, Churchward-Venne TA, Phillips SM. Bigger weights may not beget bigger muscles: evidence from acute muscle protein synthetic responses after resistance exercise. *Appl Physiol Nutr Metab Physiol Appliquée Nutr Métabolisme* 2012;37(3):551– 4.
- 9. Campos GER, Luecke TJ, Wendeln HK, et al. Muscular adaptations in response to three different resistance-training regimens: specificity of repetition maximum training zones. *Eur J Appl Physiol* 2002;88(1-2):50–60.
- 10. Chaillou T, Koulmann N, Simler N, et al. Hypoxia transiently affects skeletal muscle hypertrophy in a functional overload model. *Am J Physiol Regul Integr Comp Physiol* 2012;302(5):R643–54.
- 11. Clark BC, Fernhall B, Ploutz-Snyder LL. Adaptations in human neuromuscular function following prolonged unweighting: I. Skeletal muscle contractile properties and applied ischemia efficacy. *J Appl Physiol Bethesda Md 1985* 2006;101(1):256–63.
- 12. Cook CJ, Kilduff LP, Beaven CM. Improving strength and power in trained athletes with 3 weeks of occlusion training. *Int J Sports Physiol Perform* 2014;9(1):166–72.
- 13. Counts BR, Dankel SJ, Barnett BE, et al. The influence of relative blood flow restriction pressure on muscle activation and muscle adaptation. *Muscle Nerve* 2015;

- Damas F, Phillips SM, Lixandrão ME, et al. Early resistance training-induced increases in muscle cross-sectional area are concomitant with edema-induced muscle swelling. *Eur J Appl Physiol* 2016;116(1):49–56.
- Ellefsen S, Hammarström D, Strand TA, et al. Blood flow-restricted strength training displays high functional and biological efficacy in women: a within-subject comparison with high-load strength training. *Am J Physiol Regul Integr Comp Physiol* 2015;309(7):R767–79.
- 16. Fahs CA, Loenneke JP, Thiebaud RS, et al. Muscular adaptations to fatiguing exercise with and without blood flow restriction. *Clin Physiol Funct Imaging* 2015;35(3):167–76.
- 17. Farup J, de Paoli F, Bjerg K, Riis S, Ringgard S, Vissing K. Blood flow restricted and traditional resistance training performed to fatigue produce equal muscle hypertrophy. *Scand J Med Sci Sports* 2015;
- Fitschen PJ, Kistler BM, Jeong JH, et al. Perceptual effects and efficacy of intermittent or continuous blood flow restriction resistance training. *Clin Physiol Funct Imaging* 2014;34(5):356–63.
- 19. Fry CS, Drummond MJ, Glynn EL, et al. Skeletal muscle autophagy and protein breakdown following resistance exercise are similar in younger and older adults. *J Gerontol A Biol Sci Med Sci* 2013;68(5):599–607.
- Fry CS, Glynn EL, Drummond MJ, et al. Blood flow restriction exercise stimulates mTORC1 signaling and muscle protein synthesis in older men. *J Appl Physiol Bethesda Md 1985* 2010;108(5):1199–209.
- 21. Fujita S, Abe T, Drummond MJ, et al. Blood flow restriction during low-intensity resistance exercise increases S6K1 phosphorylation and muscle protein synthesis. *J Appl Physiol Bethesda Md 1985* 2007;103(3):903–10.
- 22. Garten RS, Goldfarb A, Crabb B, Waller J. The Impact of Partial Vascular Occlusion on Oxidative Stress Markers during Resistance Exercise. *Int J Sports Med* 2015;36(7):542–9.
- 23. Goldfarb AH, Garten RS, Chee PDM, et al. Resistance exercise effects on blood glutathione status and plasma protein carbonyls: influence of partial vascular occlusion. *Eur J Appl Physiol* 2008;104(5):813–9.
- 24. Goldspink G. Changes in muscle mass and phenotype and the expression of autocrine and systemic growth factors by muscle in response to stretch and overload. *J Anat* 1999;194 (Pt 3):323–34.
- 25. Gundermann DM, Fry CS, Dickinson JM, et al. Reactive hyperemia is not responsible for stimulating muscle protein synthesis following blood flow restriction exercise. *J Appl Physiol Bethesda Md 1985* 2012;112(9):1520–8.

- 26. Gundermann DM, Walker DK, Reidy PT, et al. Activation of mTORC1 signaling and protein synthesis in human muscle following blood flow restriction exercise is inhibited by rapamycin. *Am J Physiol Endocrinol Metab* 2014;306(10):E1198–204.
- 27. Haddad F, Adams GR. Inhibition of MAP/ERK kinase prevents IGF-I-induced hypertrophy in rat muscles. *J Appl Physiol Bethesda Md 1985* 2004;96(1):203–10.
- 28. Hardie DG. AMP-activated/SNF1 protein kinases: conserved guardians of cellular energy. *Nat Rev Mol Cell Biol* 2007;8(10):774–85.
- 29. Hayot M, Rodriguez J, Vernus B, et al. Myostatin up-regulation is associated with the skeletal muscle response to hypoxic stimuli. *Mol Cell Endocrinol* 2011;332(1-2):38–47.
- 30. Henneman E, Somjen G, Carpenter DO. FUNCTIONAL SIGNIFICANCE OF CELL SIZE IN SPINAL MOTONEURONS. *J Neurophysiol* 1965;28:560–80.
- 31. Hollander DB, Reeves GV, Clavier JD, Francois MR, Thomas C, Kraemer RR. Partial occlusion during resistance exercise alters effort sense and pain. *J Strength Cond Res Natl Strength Cond Assoc* 2010;24(1):235–43.
- 32. Hubal MJ, Gordish-Dressman H, Thompson PD, et al. Variability in muscle size and strength gain after unilateral resistance training. *Med Sci Sports Exerc* 2005;37(6):964–72.
- 33. Ito N, Ruegg UT, Kudo A, Miyagoe-Suzuki Y, Takeda S 'ichi. Activation of calcium signaling through Trpv1 by nNOS and peroxynitrite as a key trigger of skeletal muscle hypertrophy. *Nat Med* 2013;19(1):101–6.
- Jacobs BL, You J-S, Frey JW, Goodman CA, Gundermann DM, Hornberger TA. Eccentric contractions increase the phosphorylation of tuberous sclerosis complex-2 (TSC2) and alter the targeting of TSC2 and the mechanistic target of rapamycin to the lysosome. *J Physiol* 2013;591(Pt 18):4611–20.
- Kacin A, Strazar K. Frequent low-load ischemic resistance exercise to failure enhances muscle oxygen delivery and endurance capacity. *Scand J Med Sci Sports* 2011;21(6):e231–41.
- Kim E, Gregg LD, Kim L, Sherk VD, Bemben MG, Bemben DA. Hormone responses to an acute bout of low intensity blood flow restricted resistance exercise in college-aged females. *J Sports Sci Med* 2014;13(1):91–6.
- Kraemer WJ, Marchitelli L, Gordon SE, et al. Hormonal and growth factor responses to heavy resistance exercise protocols. *J Appl Physiol Bethesda Md 1985* 1990;69(4):1442– 50.
- 38. Kraemer WJ, Noble BJ, Clark MJ, Culver BW. Physiologic responses to heavy-resistance exercise with very short rest periods. *Int J Sports Med* 1987;8(4):247–52.

- Kubota A, Sakuraba K, Koh S, Ogura Y, Tamura Y. Blood flow restriction by low compressive force prevents disuse muscular weakness. *J Sci Med Sport Sports Med Aust* 2011;14(2):95–9.
- 40. Kubota A, Sakuraba K, Sawaki K, Sumide T, Tamura Y. Prevention of disuse muscular weakness by restriction of blood flow. *Med Sci Sports Exerc* 2008;40(3):529–34.
- Kumar V, Selby A, Rankin D, et al. Age-related differences in the dose-response relationship of muscle protein synthesis to resistance exercise in young and old men. J Physiol 2009;587(Pt 1):211–7.
- 42. Laurentino GC, Loenneke JP, Teixeira EL, Nakajima E, Iared W, Tricoli V. The Effect of Cuff Width on Muscle Adaptations after Blood Flow Restriction Training. *Med Sci Sports Exerc* 2015; doi: 10.1249/MSS.0000000000833.
- Laurentino GC, Ugrinowitsch C, Roschel H, et al. Strength training with blood flow restriction diminishes myostatin gene expression. *Med Sci Sports Exerc* 2012;44(3):406– 12.
- 44. Laurentino G, Ugrinowitsch C, Aihara AY, et al. Effects of strength training and vascular occlusion. *Int J Sports Med* 2008;29(8):664–7.
- 45. Loenneke JP, Abe T, Wilson JM, et al. Blood flow restriction: an evidence based progressive model (Review). *Acta Physiol Hung* 2012;99(3):235–50.
- Loenneke JP, Allen KM, Mouser JG, et al. Blood flow restriction in the upper and lower limbs is predicted by limb circumference and systolic blood pressure. *Eur J Appl Physiol* 2015;115(2):397–405.
- 47. Loenneke JP, Fahs CA, Rossow LM, et al. Effects of cuff width on arterial occlusion: implications for blood flow restricted exercise. *Eur J Appl Physiol* 2012;112(8):2903–12.
- 48. Loenneke JP, Fahs CA, Thiebaud RS, et al. The acute muscle swelling effects of blood flow restriction. *Acta Physiol Hung* 2012;99(4):400–10.
- Loenneke JP, Kim D, Fahs CA, et al. Effects of exercise with and without different degrees of blood flow restriction on torque and muscle activation. *Muscle Nerve* 2015;51(5):713–21.
- 50. Loenneke JP, Thiebaud RS, Abe T. Does blood flow restriction result in skeletal muscle damage? A critical review of available evidence. *Scand J Med Sci Sports* 2014;24(6):e415–22.
- Loenneke JP, Thiebaud RS, Fahs CA, Rossow LM, Abe T, Bemben MG. Blood flow restriction does not result in prolonged decrements in torque. *Eur J Appl Physiol* 2013;113(4):923–31.

- 52. Loenneke JP, Thrower AD, Balapur A, Barnes JT, Pujol TJ. Blood flow-restricted walking does not result in an accumulation of metabolites. *Clin Physiol Funct Imaging* 2012;32(1):80–2.
- 53. Loenneke JP, Wilson GJ, Wilson JM. A mechanistic approach to blood flow occlusion. *Int J Sports Med* 2010;31(1):1–4.
- 54. Loenneke JP, Wilson JM, Marín PJ, Zourdos MC, Bemben MG. Low intensity blood flow restriction training: a meta-analysis. *Eur J Appl Physiol* 2012;112(5):1849–59.
- 55. Lowery RP, Joy JM, Loenneke JP, et al. Practical blood flow restriction training increases muscle hypertrophy during a periodized resistance training programme. *Clin Physiol Funct Imaging* 2014;34(4):317–21.
- 56. Low SY, Rennie MJ, Taylor PM. Signaling elements involved in amino acid transport responses to altered muscle cell volume. *FASEB J Off Publ Fed Am Soc Exp Biol* 1997;11(13):1111–7.
- 57. Luebbers PE, Fry AC, Kriley LM, Butler MS. The effects of a 7-week practical blood flow restriction program on well-trained collegiate athletes. *J Strength Cond Res Natl Strength Cond Assoc* 2014;28(8):2270–80.
- 58. Madarame H, Sasaki K, Ishii N. Endocrine responses to upper- and lower-limb resistance exercises with blood flow restriction. *Acta Physiol Hung* 2010;97(2):192–200.
- 59. Marcotte GR, West DWD, Baar K. The molecular basis for load-induced skeletal muscle hypertrophy. *Calcif Tissue Int* 2015;96(3):196–210.
- 60. Mason S, Wadley GD. Skeletal muscle reactive oxygen species: a target of good cop/bad cop for exercise and disease. *Redox Rep Commun Free Radic Res* 2014;19(3):97–106.
- 61. Mattar MA, Gualano B, Perandini LA, et al. Safety and possible effects of low-intensity resistance training associated with partial blood flow restriction in polymyositis and dermatomyositis. *Arthritis Res Ther* 2014;16(5):473.
- 62. Mauro A. Satellite cell of skeletal muscle fibers. J Biophys Biochem Cytol 1961;9:493-5.
- 63. McBride JM, Blaak JB, Triplett-McBride T. Effect of resistance exercise volume and complexity on EMG, strength, and regional body composition. *Eur J Appl Physiol* 2003;90(5-6):626–32.
- 64. McCarthy JJ, Mula J, Miyazaki M, et al. Effective fiber hypertrophy in satellite celldepleted skeletal muscle. *Dev Camb Engl* 2011;138(17):3657–66.
- 65. Merry TL, Ristow M. Do antioxidant supplements interfere with skeletal muscle adaptation to exercise training? *J Physiol* 2015;

- 66. Mitchell CJ, Churchward-Venne TA, West DDW, et al. Resistance exercise load does not determine training-mediated hypertrophic gains in young men [Internet]. *J Appl Physiol* 2012; [cited 2015 Aug 12] Available from: http://jap.physiology.org/content/early/2012/04/12/japplphysiol.00307.2012
- 67. Moritani T, Sherman WM, Shibata M, Matsumoto T, Shinohara M. Oxygen availability and motor unit activity in humans. *Eur J Appl Physiol* 1992;64(6):552–6.
- 68. Morton RW, McGlory C, Phillips SM. Nutritional interventions to augment resistance training-induced skeletal muscle hypertrophy. *Front Physiol* 2015;6:245.
- 69. Motykie GD, Zebala LP, Caprini JA, et al. A guide to venous thromboembolism risk factor assessment. *J Thromb Thrombolysis* 2000;9(3):253–62.
- Nielsen JL, Aagaard P, Bech RD, et al. Proliferation of myogenic stem cells in human skeletal muscle in response to low-load resistance training with blood flow restriction. J Physiol 2012;590(Pt 17):4351–61.
- 71. Ohta H, Kurosawa H, Ikeda H, Iwase Y, Satou N, Nakamura S. Low-load resistance muscular training with moderate restriction of blood flow after anterior cruciate ligament reconstruction. *Acta Orthop Scand* 2003;74(1):62–8.
- 72. Oishi Y, Tsukamoto H, Yokokawa T, et al. Mixed lactate and caffeine compound increases satellite cell activity and anabolic signals for muscle hypertrophy. *J Appl Physiol Bethesda Md 1985* 2015;118(6):742–9.
- 73. Okamoto T, Masuhara M, Ikuta K. Differences of muscle oxygenation during eccentric and concentric contraction. *Isokinet Exerc Sci* 2006;14(3):207–12.
- 74. Ozaki H, Loenneke JP, Buckner SL, Abe T. Muscle growth across a variety of exercise modalities and intensities: Contributions of mechanical and metabolic stimuli [Internet]. *Med Hypotheses* [date unknown];0(0) [cited 2016 Jan 10] Available from: http://www.medical-hypotheses.com/article/S0306987716000098/abstract
- 75. Ozaki H, Sakamaki M, Yasuda T, et al. Increases in thigh muscle volume and strength by walk training with leg blood flow reduction in older participants. *J Gerontol A Biol Sci Med Sci* 2011;66(3):257–63.
- 76. Patterson SD, Ferguson RA. Increase in calf post-occlusive blood flow and strength following short-term resistance exercise training with blood flow restriction in young women. *Eur J Appl Physiol* 2010;108(5):1025–33.
- Patterson SD, Leggate M, Nimmo MA, Ferguson RA. Circulating hormone and cytokine response to low-load resistance training with blood flow restriction in older men. *Eur J Appl Physiol* 2013;113(3):713–9.
- 78. Phillips SM. A brief review of critical processes in exercise-induced muscular hypertrophy. *Sports Med Auckl NZ* 2014;44 Suppl 1:S71–7.

- Phillips SM, Tipton KD, Aarsland A, Wolf SE, Wolfe RR. Mixed muscle protein synthesis and breakdown after resistance exercise in humans. *Am J Physiol* 1997;273(1 Pt 1):E99– 107.
- 80. Powers SK, Ji LL, Kavazis AN, Jackson MJ. Reactive oxygen species: impact on skeletal muscle. *Compr Physiol* 2011;1(2):941–69.
- Reeves GV, Kraemer RR, Hollander DB, et al. Comparison of hormone responses following light resistance exercise with partial vascular occlusion and moderately difficult resistance exercise without occlusion. *J Appl Physiol Bethesda Md 1985* 2006;101(6):1616–22.
- 82. Sakamaki M, G Bemben M, Abe T. Legs and trunk muscle hypertrophy following walk training with restricted leg muscle blood flow. *J Sports Sci Med* 2011;10(2):338–40.
- 83. Sandri M. Signaling in muscle atrophy and hypertrophy. *Physiol Bethesda Md* 2008;23:160–70.
- 84. Sato Y. The history and future of KAATSU Training. *Int J KAATSU Train Res* 2005;1(1):1–5.
- 85. Scott BR, Dascombe BJ, Delaney JA, et al. The validity and reliability of a customized rigid supportive harness during Smith machine back squat exercise. *J Strength Cond Res Natl Strength Cond Assoc* 2014;28(3):636–42.
- 86. Suga T, Okita K, Takada S, et al. Effect of multiple set on intramuscular metabolic stress during low-intensity resistance exercise with blood flow restriction. *Eur J Appl Physiol* 2012;112(11):3915–20.
- 87. Takano H, Morita T, Iida H, et al. Hemodynamic and hormonal responses to a short-term low-intensity resistance exercise with the reduction of muscle blood flow. *Eur J Appl Physiol* 2005;95(1):65–73.
- 88. Takarada Y, Nakamura Y, Aruga S, Onda T, Miyazaki S, Ishii N. Rapid increase in plasma growth hormone after low-intensity resistance exercise with vascular occlusion. *J Appl Physiol Bethesda Md 1985* 2000;88(1):61–5.
- Timmerman KL, Lee JL, Dreyer HC, et al. Insulin stimulates human skeletal muscle protein synthesis via an indirect mechanism involving endothelial-dependent vasodilation and mammalian target of rapamycin complex 1 signaling. *J Clin Endocrinol Metab* 2010;95(8):3848–57.
- 90. Toigo M, Boutellier U. New fundamental resistance exercise determinants of molecular and cellular muscle adaptations. *Eur J Appl Physiol* 2006;97(6):643–63.
- 91. Van der Meer SFT, Jaspers RT, Degens H. Is the myonuclear domain size fixed? J Musculoskelet Neuronal Interact 2011;11(4):286–97.

- 92. Vechin FC, Libardi CA, Conceição MS, et al. Comparisons between low-intensity resistance training with blood flow restriction and high-intensity resistance training on quadriceps muscle mass and strength in elderly. J Strength Cond Res Natl Strength Cond Assoc 2015;29(4):1071–6.
- Vikne H, Refsnes PE, Ekmark M, Medbø JI, Gundersen V, Gundersen K. Muscular performance after concentric and eccentric exercise in trained men. *Med Sci Sports Exerc* 2006;38(10):1770–81.
- 94. Villanueva MG, Lane CJ, Schroeder ET. Short rest interval lengths between sets optimally enhance body composition and performance with 8 weeks of strength resistance training in older men. *Eur J Appl Physiol* 2015;115(2):295–308.
- 95. Weir JP. Quantifying test-retest reliability using the intraclass correlation coefficient and the SEM. J Strength Cond Res Natl Strength Cond Assoc 2005;19(1):231–40.
- 96. West DWD, Burd NA, Tang JE, et al. Elevations in ostensibly anabolic hormones with resistance exercise enhance neither training-induced muscle hypertrophy nor strength of the elbow flexors. *J Appl Physiol Bethesda Md 1985* 2010;108(1):60–7.
- 97. Wilson JM, Lowery RP, Joy JM, Loenneke JP, Naimo MA. Practical blood flow restriction training increases acute determinants of hypertrophy without increasing indices of muscle damage. *J Strength Cond Res Natl Strength Cond Assoc* 2013;27(11):3068–75.
- 98. Yasuda T, Brechue WF, Fujita T, Sato Y, Abe T. Muscle activation during low-intensity muscle contractions with varying levels of external limb compression. *J Sports Sci Med* 2008;7(4):467–74.
- 99. Yasuda T, Fukumura K, Fukuda T, et al. Effects of low-intensity, elastic band resistance exercise combined with blood flow restriction on muscle activation. *Scand J Med Sci Sports* 2014;24(1):55–61.
- 100. Yasuda T, Loenneke JP, Thiebaud RS, Abe T. Effects of blood flow restricted low-intensity concentric or eccentric training on muscle size and strength. *PloS One* 2012;7(12):e52843.
- 101. Yasuda T, Ogasawara R, Sakamaki M, Ozaki H, Sato Y, Abe T. Combined effects of lowintensity blood flow restriction training and high-intensity resistance training on muscle strength and size. *Eur J Appl Physiol* 2011;111(10):2525–33.
- 102. Zanou N, Gailly P. Skeletal muscle hypertrophy and regeneration: interplay between the myogenic regulatory factors (MRFs) and insulin-like growth factors (IGFs) pathways. *Cell Mol Life Sci CMLS* 2013;70(21):4117–30.

VITA

Scott Dankel, BA 224 James Circle Oxford, MS Phone: (732) 610-6308 Email: sdankel@go.olemiss.edu

EDUCATION

- 2012 B.A. Health and Physical Education Rowan University
- 2016 M.S. Exercise Science The University of Mississippi

PROFESSIONAL EXPERIENCE

- 2012 Health and Physical Education Teaching Assistant at Deptford High School; Deptford, NJ
- 2011 Health and Physical Education Teaching Assistant at Lake Tract Elementary School; Deptford, NJ
- 2016 Graduate Teaching Assistant at The University of Mississippi

TEACHING EXPERIENCE

- 1) Exercise Physiology Lab (1 section): Summer 2015 (1 section)
- 2) Exercise Prescription Lab (5 sections): Summer 2015 (2 sections), Fall 2015 (1 section), Spring 2016 (2 sections)
- 3) Personal and Community Health (1 section): Fall 2015 (1 section)
- 4) First Aid and CPR (1 section): Spring 2016 (1 section)
- 5) Tennis (1 section): Fall 2014 (1 section)
- 6) Weight Lifting (9 sections): Fall 2014 (3 sections), Spring 2015 (4 sections), Fall 2015 (1 section), Spring 2016 (1 section)
- 7) Jogging (2 sections): Fall 2014 (1 section), Spring 2015 (1 section

GRANTS

- 1) Loenneke JP, Dankel SJ (2015). The effects of metabolic stress on muscle size and strength. The Biolayne Foundation. \$10,000 (FUNDED). Intellectually contributed to to the conception of the study and assisted with drafting the methods and grant application.
- 2) Loenneke JP (2015). Can muscle growth occur through resistance training with no external load? American College of Sports Medicine \$ (Under Review). Assisted with drafting the methods and grant application.

PEER-REVIEWED PUBLICATIONS

- Abe T, Counts BR, Barnett BE, Dankel SJ, Lee K, and Loenneke JP (2015). Associations between Handgrip Strength and Ultrasound-Measured Muscle Thickness of the Hand and Forearm in Young Men and Women. Ultrasound in medicine & biology.
- 2) Barnett BE, **Dankel SJ**, Counts BR, Nooe AL, Abe T, and Loenneke JP (2015). Blood flow occlusion pressure at rest and immediately after a bout of low load exercise. Clinical physiology and functional imaging.
- 3) Buckner SL, Abe T, Counts BR, **Dankel SJ**, Barnett BE, and Loenneke JP (2015). Muscle and fat mapping of the trunk: a case study. Journal of Ultrasound.
- 4) Counts BR, **Dankel SJ**, Barnett BE, Kim D, Mouser JG, Allen KM, Thiebaud RS, Abe T, Bemben MG, and Loenneke, JP (2015). The influence of relative blood flow restriction pressure on muscle activation and muscle adaptation. Muscle & nerve.
- 5) **Dankel SJ,** Loenneke JP, and Loprinzi PD (2015). Participation in muscle strengthening activities as an alternative method for the prevention of multimorbidity. Preventive Medicine.
- 6) **Dankel SJ,** Loenneke JP, and Loprinzi PD (2015). The impact of overweight/obesity duration on the association between physical activity and cardiovascular disease risk: an application of the "fat but fit" paradigm. International Journal of Cardiology.
- 7) **Dankel SJ,** Loenneke JP, and Loprinzi PD (2015). Physical activity and diet on quality of life and mortality: The importance of meeting one specific or both behaviors. International Journal of Cardiology.
- 8) **Dankel SJ,** Loenneke JP, and Loprinzi PD (2015). The Effects of Blood Flow Restriction on Upper-Body Musculature Located Distal and Proximal to Applied Pressure. Sports Medicine.
- 9) Dankel SJ, Loenneke JP, and Loprinzi PD (2015). Does the fat-but-fit paradigm hold true for all-cause mortality when considering the duration of overweight/obesity? Analyzing the WATCH (Weight, Activity and Time Contributes to Health) Paradigm. Preventive Medicine.
- 10) Dankel SJ, Loenneke JP, and Loprinzi PD (2015). Determining the Importance of Meeting Muscle-Strengthening Activity Guidelines: Is the Behavior or the Outcome of the Behavior (Strength) a More Important Determinant of All-cause Mortality?. Mayo Clinic Proceedings.

- 11) Dankel SJ, Loenneke JP, and Loprinzi PD (2016). Combined Associations of Muscle-Strengthening Activities and Accelerometer-Assessed Physical Activity on Multimorbidity: Findings From NHANES. American Journal of Health Promotion.
- 12) **Dankel SJ,** Loenneke JP, and Loprinzi PD (2016). Mild depressive symptoms amongst Americans in relation to physical activity, current overweight/obesity, and self-reported history of overweight/obesity. International Journal of Behavioral Medicine.
- 13) Dankel SJ, Loenneke JP, and Loprinzi PD (2016). The WATCH (Weight Activity and Time Contributes to Health) paradigm and quality of life: the impact of overweight/obesity duration on the association between physical activity and healthrelated quality of life. The International Journal of Clinical Practice.
- 14) Jessee MB, Buckner SL, **Dankel SJ**, Counts BR, Abe T, and Loenneke JP (2016). The Influence of Cuff Width, Sex, and Race on Arterial Occlusion: Implications for Blood Flow Restriction Research. Sports Medicine.

SCIENTIFIC ABSTRACTS/ORAL PRESENTATIONS

- Barnett BE, Dankel SJ, Counts BR, Nooe AL, Abe T, Loenneke JP. Predictors of standing upper body arterial occlusion: implications for blood flow restriction research. ACSM National Conference, May 2015, San Diego, California.
- 2) Counts BR, **Dankel SJ**, Barnett BE, Abe T, Loenneke JP. High relative pressures do not augment changes in early phase muscular adaptations during blood flow restricted exercise. ACSM National Conference, May 2015, San Diego, California
- Dankel SJ, Barnett BE, Counts BR, Nooe AL, Abe T, Loenneke JP. Blood flow occlusion pressure at rest and immediately after a bout of low load exercise. ACSM National Conference, May 2015, San Diego, California.
- 4) Counts BR, Buckner SL, **Dankel SJ**, Jessee MB, Mattocks KT, Mouser JG, Laurentino GC, and Loenneke JP. The Acute Response to No Load Exercise: Is it Sufficient? ACSM National Conference, May 2016, Boston, Massachusetts.
- 5) Barnett BE, Buckner SL, **Dankel SJ**, Counts BR, Jessee MB, Mouser JG, Halliday TM and Loenneke JP. Circadian Rhythms in Blood Glucose and Blood Pressure: Are they Reproducible? ACSM National Conference, May 2016, Boston, Massachusetts.
- 6) Mouser JG, Buckner SL, Counts BR, **Dankel SJ**, Jessee MB, Mattocks KT, Laurentino GC, and Loenneke JP. Venous versus Arterial Blood Flow Restriction: The Impact of Cuff Width. ACSM National Conference, May 2016, Boston, Massachusetts.
- Ingram JW, Buckner SL, Dankel SJ, Counts BR, Mouser JG, Abe T, Laurentino GC, and Loenneke JP. The influence of time on determining blood flow restriction pressure. ACSM National Conference, May 2016, Boston, Massachusetts.
- Mattocks KT, Buckner SL, Dankel SJ, Counts BR, Jessee MB, Mouser JG, Laurentinio GC, Abe T, and Loenneke JP. The Influence of Cuff Material on the Blood Flow Restriction Stimulus in the Upper Body. ACSM National Conference, May 2016, Boston, Massachusetts.
- 9) Laurentino GC, Mouser JG, Buckner SL, Counts BR, **Dankel SJ**, Jessee MB, Mattocks KT, Loenneke JP, Tricoli V. The influence of cuff width on regional muscle growth:

Implications for Blood Flow Restriction Training. ACSM National Conference, May 2016, Boston, Massachusetts.

- 10) Jessee MB, Buckner SL, **Dankel SJ**, Counts BR, Abe T, and Loenneke JP. The Influence of Cuff Width and Sex on Arterial Occlusion: Implications for Blood Flow Restriction Research. ACSM National Conference, May 2016, Boston, Massachusetts.
- 11) Loenneke JP, Buckner SL, Dankel SJ, Jessee MB, Counts BR, Mouser JG, Mattocks KT, Laurentino GC, and Abe T. The Influence of Cuff Material on the Acute Muscular Response to Blood Flow Restricted Exercise in the Upper Body. ACSM National Conference, May 2016, Boston, Massachusetts.
- 12) Buckner SL, Dankel SJ, Counts BR, Barnett BE, Jessee MB, Mouser JG, Halliday TM, and Loenneke JP. The Influence of Circadian Rhythms on Upper Body Isometric Strength, Muscle Thickness and Body Temperature. ACSM National Conference, May 2016, Boston, Massachusetts.
- 13) Dankel SJ, Counts BR, Barnett BE, Buckner SL, Abe T, Zourdos MC, and Loenneke JP. Muscle adaptation to 21 Straight Days of Elbow Flexor Exercise in Trained Individuals. ACSM National Conference, May 2016, Boston, Massachusetts.

HONORS AND AWARDS

2014 J. Robert Blackburn Graduate Award in Exercise Science

MENTORSHIP

Jeremy Loenneke, PhD The University of Mississippi (2014 – 2016)